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(54) PHARMACEUTICAL COMPOSITION FOR TREATMENT AND/OR PREVENTION OF CANCER

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(57) ABSTRACT

This invention provides an antibody targeting a cancer antigenic protein specifically expressed on the surface of cancer cells and use thereof in a therapeutic and/or preventive agent for cancer. Specifically, this invention provides an antibody or a fragment thereof which has immunological reactivity with a partial CAPRIN-1 polypeptide consisting of the amino acid sequence set forth in SEQ ID NO: 5 or an amino acid sequence having 80% or higher sequence identity to the amino acid sequence, and a pharmaceutical composition for treatment and/or prevention of cancer, comprising the antibody or fragment thereof as an active ingredient.

20 Claims, No Drawings

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PHARMACEUTICAL COMPOSITION FOR TREATMENT AND/OR PREVENTION OF **CANCER**

SEQUENCE LISTING

The text file titled Revised_Seq_List_09242015_2.txt of size 50 KB created Sep. 24, 2015, filed herewith, is hereby incorporated by reference.

TECHNICAL FIELD

The present invention relates to novel use of an antibody against CAPRIN-1 or a fragment thereof in a drug such as a therapeutic and/or preventive agent for cancer.

BACKGROUND ART

Cancer is the leading cause of death. This disease is currently treated principally by surgical therapy in combination 20 with radiation therapy and/or chemotherapy. In spite of recent development of novel surgical techniques or discovery of novel anticancer agents, the existing treatment of cancer has an insufficiently improved outcome, except for some cancer types. With recent advances of molecular biology or cancer 25 immunology, antibodies that specifically react with cancer, cancer antigens that are recognized by cytotoxic T cells, genes encoding such cancer antigens, and the like have been identified, raising expectations on specific cancer therapy targeting the cancer antigens (Non Patent Literature 1).

For reducing the adverse reaction of cancer therapy, it is desired that peptides, polypeptides, or proteins recognized as antigens of the cancer should rarely exist in normal cells and specifically exist in cancer cells. In 1991, Boon et al. (Ludwig Institute for Cancer Research, Belgium) isolated a human 35 melanoma antigen MAGE1 recognized by CD8-positive T cells by a cDNA expression cloning method using autologous cancer cell lines and cancer-reactive T cells (Non Patent Literature 2). Then, a SEREX (serological identification of antigens by recombinant expression cloning) method has been 40 reported, which adopts a gene expression cloning approach to identify tumor antigens recognized by antibodies produced in response to autologous cancer in vivo in a cancer patient (Non Patent Literature 3 and Patent Literature 1). According to this method, some cancer antigens that are rarely expressed in 45 normal cells and are specifically expressed in cancer have been isolated (Non Patent Literatures 4 to 9). In addition, cell therapy using immunocytes that specifically react with cancer antigens or cancer-specific immunotherapy using vaccines or the like comprising cancer antigens is under clinical trial 50 targeting some of the isolated cancer antigens.

In recent years, various antibody drugs for cancer treatment targeting antigenic proteins on cancer cells have emerged in the world. These drugs have received attention agents. A large majority of antigenic proteins targeted by the drugs, however, are also expressed in normal cells. As a result of administering the antibodies, cancer cells as well as normal cells expressing the antigens are damaged, disadvantageously resulting in adverse reaction. Thus, if cancer antigens specifi- 60 cally expressed on the surface of cancer cells can be identified and antibodies targeting the antigens can be used as drugs, these antibody drugs can be expected to achieve treatment with less adverse reaction.

Cytoplasmic- and proliferation-associated protein 1 (CA- 65 PRIN-1) has been known as an intracellular protein that is expressed upon activation or cell division of resting normal

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cells and forms cytoplasmic stress granules with intracellular RNAs to participate in the regulation of transport and translation of mRNAs. This protein has been found to be specifically expressed on the surface of cancer cells and is under study as a target of antibody drugs for cancer treatment (Patent Literature 2).

CITATION LIST

Patent Literature

Patent Literature 1: U.S. Pat. No. 5,698,396 Patent Literature 2: WO2010/016526

Non Patent Literature

Non Patent Literature 1: Tsuyoshi Akiyoshi, "Japanese Journal of Cancer and Chemotherapy", 1997, Vol. 24, p. 55-519 (Japanese Journal of Cancer and Chemotherapy Publishers Inc., Japan)

Non Patent Literature 2: Bruggen P. et al., Science, 254: 1643-1647 (1991)

Non Patent Literature 3: Proc. Natl. Acad. Sci. USA, 92: 11810-11813 (1995)

Non Patent Literature 4: Int. J. Cancer, 72: 965-971 (1997) Non Patent Literature 5: Cancer Res., 58: 1034-1041 (1998) Non Patent Literature 6: Int. J. Cancer, 29: 652-65.8 (1998) Non Patent Literature 7: Int. J. Oncol., 14: 703-708 (1999)

Non Patent Literature 8: Cancer Res., 56: 4766-4772 (1996) Non Patent Literature 9: Hum. Mol. Genet 6: 33-39, 1997

SUMMARY OF INVENTION

Technical Problem

An object of the present invention is to produce an antibody that targets CAPRIN-1 specifically expressed on the surface of cancer cells and is superior in antitumor activity to conventional antibodies, and to provide use thereof as a therapeutic and/or preventive agent for cancer.

Solution to Problem

Features of the present invention are as follows:

The present invention provides an antibody or a fragment thereof which has immunological reactivity with a partial CAPRIN-1 polypeptide having the amino acid sequence set forth in SEQ ID NO: 5 or an amino acid sequence having 80% or higher sequence identity to the amino acid sequence, and a pharmaceutical composition for treatment and/or prevention of cancer, comprising the antibody or fragment thereof as an active ingredient.

In the above embodiment, the cancer is breast cancer, kidbecause of their certain efficacy as cancer-specific therapeutic 55 ney cancer, pancreatic cancer, large intestinal cancer, lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary bladder cancer, esophageal cancer, leukemia, lymphoma, fibrosarcoma, mastocytoma, or melanoma.

> In one embodiment, the antibody is a monoclonal antibody or a polyclonal antibody.

> In another embodiment, the antibody is a human antibody, a humanized antibody, a chimeric antibody, a single-chain antibody, or a multispecific antibody (e.g., a bispecific antibody).

> The present specification encompasses the contents described in the specification and/or drawings of Japanese

Patent Application No. 2012-035491 on which the priority of the present application is based.

Advantageous Effects of Invention

The antibody against CAPRIN-1 according to the present invention damages cancer cells. Thus, the antibody against CAPRIN-1 is useful in the treatment or prevention of cancer.

DESCRIPTION OF EMBODIMENTS

The antibody according to the present invention is an antibody that recognizes and binds to a predetermined partial polypeptide of CAPRIN-1 and has antitumor activity. More specifically, the antibody according to the present invention is an antibody that recognizes (i.e., has immunological reactivity with) a partial polypeptide of a CAPRIN-1 protein (partial CAPRIN-1 polypeptide) consisting of the amino acid sequence set forth in SEQ ID NO 5 or an amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence. In the present invention, this antibody has been shown to exhibit antitumor activity. The present invention relates to all antibodies that bind to fragments of CAPRIN-1 proteins as described above 25 and exhibit antitumor activity.

The antibody against CAPRIN-1 according to the present invention may be any type of antibody that can exert antitumor activity and includes, for example, recombinant antibodies (e.g., synthetic antibodies, multispecific antibodies (e.g., 30 bispecific antibodies), humanized antibodies, chimeric antibodies, and single-chain antibodies (scFv)), human antibodies, and their antibody fragments (e.g., Fab, F(ab')₂, and Fv). These antibodies and fragments thereof can be prepared by methods generally known to those skilled in the art. Desir- 35 ably, the antibody according to the present invention has immunological reactivity with a CAPRIN-1 protein or a partial polypeptide thereof, i.e., binds to (preferably, specifically binds to) the CAPRIN-1 protein through antigen-antibody reaction. In this context, the phrase "specifically binding to 40 the CAPRIN-1 protein" means that the antibody specifically binds to the CAPRIN-1 protein without substantially binding to other proteins. The antibody according to the present invention is preferably a monoclonal antibody and however, may be a polyclonal antibody as long as homogeneous anti- 45 bodies can be stably produced. When a subject is a human, a human antibody or a humanized antibody is desirable for avoiding or suppressing rejection.

The antibody against a CAPRIN-1 polypeptide according to the present invention can be evaluated for its antitumor 50 activity, as described later, by examining in vivo the inhibition of tumor growth in a cancer-bearing animal or by examining ex vivo the presence or absence of immunocyte- or complement-mediated cytotoxic activity exhibited by the antibody against tumor cells expressing the polypeptide.

The subject to receive the treatment and/or prevention of cancer according to the present invention is a mammal such as a human, a pet animal, livestock, or a sport animal and is preferably a human.

Hereinafter, the present invention will be described in more 60 detail.

<Preparation of Antigen for Antibody Preparation>

Proteins or fragments thereof used as sensitizing antigens for obtaining the antibody against CAPRIN-1 according to the present invention are not limited by animal species serving as their origins, including humans, dogs, cats, cattle, horses, mice, rats, and chickens. The proteins or the frag-

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ments thereof, however, are preferably selected in view of compatibility with parent cells for use in cell fusion. In general, mammal-derived proteins are preferred. Particularly, human-derived proteins are preferred. For example, when CAPRIN-1 is human CAPRIN-1, human CAPRIN-1 proteins, partial peptides thereof, or cells expressing human CAPRIN-1 can be used.

The nucleotide sequences and amino acid sequences of human CAPRIN-1 and homologs thereof can be obtained, for example, by making an access to GenBank (NCBI, USA) and using an algorithm such as BLAST or FASTA (Karlin and Altschul, Proc. Natl. Acad. Sci. USA, 90: 5873-5877, 1993; and Altschul et al., Nucleic Acids Res. 25: 3389-3402, 1997).

In the present invention, with reference to the nucleotide sequence (SEQ ID NO: 1 or 3) or amino acid sequence (SEQ ID NO: 2 or 4) of human CAPRIN-1, the target CAPRIN-1 is a nucleic acid or a protein consisting of a sequence having 70% to 100%, preferably 80% to 100%, more preferably 90% to 100%, further preferably 95% to 100%, for example, 97% to 100%, 98% to 100%, 99% to 100%, or 99.5% to 100% sequence identity to the nucleotide sequence or amino acid sequence of the ORF or mature portion of the reference (note that the amino acid sequences of SEQ ID NO: 2 and SEQ ID NO: 4 compared with each other differ in amino acid residues at and following position 690). In this context, the term "% sequence identity" means a percentage (%) of the number of identical amino acids (or bases) to the total number (including the number of gaps) of amino acids (or bases) when two sequences are aligned such that the maximum degree of similarity or identity can be achieved with or without introduced

A fragment that comprises an epitope (or an antigenic determinant), which is the smallest unit recognized by the antibody, and has a length ranging from the amino acid length of the epitope to less than the full-length of the CAPRIN-1 protein can be used as a CAPRIN-1 protein fragment. The epitope refers to a polypeptide fragment having antigenicity or immunogenicity in mammals, preferably humans. Its smallest unit consists of approximately 7 to 12 amino acids, for example, 8 to 11 amino acids. The CAPRIN-1 protein fragment for use in the preparation of the antibody according to the present invention is preferably a fragment that is recognized by the antibody of the present invention and comprises the amino acid sequence set forth in SEQ ID NO: 5 (which corresponds to a sequence from positions 513 to 528 in the amino acid sequence of SEQ ID NO: 2 or 4) or an amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence, or comprises at least an epitope consisting of approximately 7 to 12 consecutive amino acids, for example, 8 to 11 consecutive amino acids in any of these amino acid sequences.

The above human CAPRIN-1 proteins and polypeptide fragments comprising partial peptides thereof can be synthesized according to chemical synthesis methods, for example, Fmoc (fluorenylmethyloxycarbonyl) and tBoc (t-butyloxycarbonyl) methods (Seikagaku Jikken Koza (Biochemical Experimentation Course in English) 1, the Japanese Biochemical Society ed., Protein Chemistry IV, Chemical Modification and Peptide Synthesis, Tokyo Kagaku Dojin Co., Ltd. (Japan), 1981). Also, these polypeptides can be synthesized by routine methods using various commercially available peptide synthesizers.

Alternatively, polynucleotides encoding the polypeptides may be prepared using genetic engineering approaches known in the art (Sambrook et al., Molecular Cloning, the 2nd edition, Current Protocols in Molecular Biology (1989), Cold

Spring Harbor Laboratory Press; Ausubel et al., Short Protocols in Molecular Biology, the 3rd edition, A compendium of Methods from Current Protocols in Molecular Biology (1995), John Wiley & Sons; etc.) and incorporated into expression vectors, which are then introduced into host cells so that the host cells produce the polypeptides. In this way, the human CAPRIN-1 proteins of interest or the polypeptide fragments thereof can be obtained.

The polynucleotides encoding the polypeptides can be readily prepared by genetic engineering approaches known in 10 the art or routine methods using commercially available nucleic acid synthesizers. For example, a DNA comprising the nucleotide sequence of a human CAPRIN-1 gene can be prepared by PCR using a human chromosomal DNA or cDNA library as a template and a pair of primers designed so 15 as to be capable of amplifying the nucleotide sequence. Reaction conditions for this PCR can be appropriately determined. Examples of the conditions can include, but not limited to, 30 cycles each involving reaction steps of 94° C. for 30 seconds (denaturation), 55° C. for 30 seconds to 1 minute (annealing), 20 and 72° C. for 2 minutes (elongation) using thermostable DNA polymerase (e.g., Taq polymerase or Pfu polymerase) and a Mg2+-containing PCR buffer, followed by reaction at 72° C. for 7 minutes. The PCR approach, conditions, etc. are described in, for example, Ausubel et al., Short Protocols in 25 Molecular Biology, the 3rd edition, A Compendium of Methods from Current Protocols in Molecular Biology (1995), John Wiley & Sons (particularly, Chapter 15).

Also, appropriate probes or primers can be prepared on the basis of information about the nucleotide sequences of 30 CAPRIN-1 genes and the amino acid sequences of CAPRIN-1 proteins, and used in the screening of, for example, a human cDNA library, to isolate the desired DNA. Preferably, such a cDNA library is produced from cells, organs, or tissues expressing CAPRIN-1 proteins. Examples 35 of such cells or tissues include cells or tissues derived from the testis or from cancers or tumors such as testis, leukemia, breast cancer, lymphoma, brain tumor, lung cancer, pancreatic cancer, and large intestinal cancer. These operations, including the preparation of probes or primers, the construc- 40 tion of a cDNA library, the screening of the cDNA library, and the cloning of the gene of interest, are known to those skilled in the art and can be performed according to methods described in, for example, Sambrook et al., Molecular Cloning, the 2nd edition, Current Protocols in Molecular Biology 45 (1989), and Ausubel et al. (ibid.). DNAs encoding the human CAPRIN-1 proteins and the partial peptides thereof can be obtained from the DNA thus obtained.

The host cells to receive the expression vectors may be any cell capable of expressing the above polypeptides. Examples 50 of prokaryotic cells include, but not limited to, *E. coli*. Examples of eukaryotic cells include, but not limited to: mammalian cells such as monkey kidney cells COS 1 and Chinese hamster ovary cells CHO; a human embryonic kidney cell line HEK293; a mouse embryonic skin cell line 55 NIH3T3; yeast cells such as budding yeast and fission yeast cells; silkworm cells; and *Xenopus* egg cells.

In the case of using prokaryotic cells as the host cells, the expression vectors used have an origin that permits replication in the prokaryotic cells, a promoter, a ribosomal binding 60 site, a multicloning site, a terminator, a drug resistance gene, an auxotrophic complementary gene, etc. Examples of expression vectors for *E. coli* can include pUC series, pBluescript II, pET expression systems, and pGEX expression systems. The DNAs encoding the above polypeptides can be 65 incorporated into such expression vectors, with which prokaryotic host cells are then transformed, followed by cul-

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ture of the obtained transformants so that the polypeptides encoded by the DNAs are expressed in the prokaryotic host cells. In this respect, the polypeptides may be expressed as fusion proteins with other proteins.

In the case of using eukaryotic cells as the host cells, expression vectors for eukaryotic cells having a promoter, a splicing region, a poly(A) addition site, etc. are used as the expression vectors. Examples of such expression vectors can include pKA1, pCDM8, pSVK3, pMSG, pSVL, pBK-CMV, pBK-RSV, EBV, pRS, pcDNA3, and pYES2 vectors. In the same way as above, the DNAs encoding the above polypeptides can be incorporated into such expression vectors, with which eukaryotic host cells are then transformed, followed by culture of the obtained transformants so that the polypeptides encoded by the DNAs are expressed in the eukaryotic host cells. In the case of using expression vectors such as pIND/ V5-His, pFLAG-CMV-2, pEGFP-N1, or pEGFP-C1, the polypeptides may be expressed as various fusion proteins tagged with, His tag (e.g., (His)₆ to (His)₁₀), FLAG tag, myc tag, HA tag, GFP, or the like.

The expression vectors can be introduced into the host cells using well known methods such as electroporation, a calcium phosphate method, a liposome method, a DEAE dextran method, microinjection, viral infection, lipofection, and binding with cell-penetrating peptides.

The polypeptide of interest can be isolated and purified from the host cells by a combination of separation operations known in the art. Examples thereof include, but not limited to, treatment with a denaturant (e.g., urea) or a surfactant, ultrasonication, enzymatic digestion, salting-out, solvent fractionation and precipitation, dialysis, centrifugation, ultrafiltration, gel filtration, SDS-PAGE, isoelectric focusing electrophoresis, ion-exchange chromatography, hydrophobic chromatography, affinity chromatography, and reverse-phase chromatography.

The antigens thus prepared can be used as sensitizing antigens as described later for producing the antibody according to the present invention.

<Structure of Antibody>

Antibodies (immunoglobulins) are usually heteromultimeric glycoproteins each comprising at least two heavy chains and two light chains. The immunoglobulins, except for IgM, are heterotetrameric glycoproteins of approximately 150 kDa each composed of two identical light (L) chains and two identical heavy (H) chains. Typically, each light chain is connected to a heavy chain via a single covalent disulfide bond, though the number of disulfide bonds between heavy chains varies among different immunoglobulin isotypes. Each of the heavy and light chains also has an intrachain disulfide bond. Each heavy chain has a variable domain (VH region) at one end, followed by a series of constant regions. Each light chain has a variable domain (VL region) at one end and has a single constant region at the other end. The light chain constant region is aligned with the first heavy chain constant region, while the light chain variable domain is aligned with the heavy chain variable domain. Particular regions called complementarity determining regions (CDRs) in the antibody variable domains exhibit specific variability and impart binding specificity to the antibody. Portions relatively conserved in the variable regions are called framework regions (FRs). The complete heavy and light chain variable domains each comprise four FRs connected via three CDRs. These three CDRs are called CDRH1, CDRH2, and CDRH3 in this order from the N-terminus of the heavy chain. Likewise, the CDRs are called CDRL1, CDRL2, and CDRL3 in the light chain. CDRH3 is most important for the binding specificity of the antibody for its antigen. In addition, CDRs

in each chain are kept close to each other by the FR regions and contribute to the formation of an antigen-binding site in the antibody, together with CDRs in the other chain. The constant regions do not directly contribute to antibody-antigen binding, but exhibit various effector functions, for example, involvement in antibody-dependent cellular cytotoxicity (ADCC), phagocytosis mediated by binding to an Fey receptor, half-life/clearance rate mediated by a neonatal Fc receptor (FcRn), and complement-dependent cytotoxicity (CDC) mediated by a C1q component in the complement cascade

<Pre><Preparation of Antibody>

The anti-CAPRIN-1 antibody according to the present invention means an antibody having immunological reactivity with a full-length CAPRIN-1 protein or a fragment thereof. Particularly, the anti-CAPRIN-1 antibody of the present invention is an antibody immunologically binding to a partial polypeptide of a CAPRIN-1 protein (partial CAPRIN-1 polypeptide) that is a peptide consisting of the 20 epitope-containing amino acid sequence set forth in SEQ ID NO: 5 or a polypeptide consisting of an amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence. Preferably, the 25 antibody of the present invention recognizes an epitope consisting of approximately 7 to 12 consecutive amino acids, for example, 8 to 11 consecutive amino acids, in the amino acid sequence set forth in SEQ ID NO: 5 or the amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence. This anti-CAPRIN-1 antibody of the present invention is capable of specifically binding to the full-length CAPRIN-1 protein. 35 The antibody of the present invention can be obtained by selecting an antibody immunologically binding to the polypeptide consisting of the amino acid sequence set forth in SEQ ID NO: 5 or the polypeptide consisting of the amino acid sequence having 80% or higher, preferably 85% or higher, 40 more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence, according to a routine method from among antibodies obtained with CAPRIN-1 proteins or fragments thereof as antigens.

In this context, the "immunological reactivity" means the 45 property of the antibody binding to the CAPRIN-1 antigen (full-length CAPRIN-1 protein or partial polypeptide thereof) in vivo. Via such binding to CAPRIN-1, the antibody of the present invention exerts the function of damaging (e.g., killing, suppressing, or regressing) tumor cells. The antibody of the present invention can damage tumor, for example, breast cancer, kidney cancer, pancreatic cancer, large intestinal cancer (e.g., colon cancer), lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary bladder cancer, esophageal cancer, leukemia, 55 lymphoma, fibrosarcoma, mastocytoma, or melanoma through binding to the CAPRIN-1 protein.

The antibody of the present invention may be any type of antibody. Examples of the type of the antibody of the present invention include monoclonal antibodies, polyclonal antibodies, synthetic antibodies, multispecific antibodies, human antibodies, humanized antibodies, chimeric antibodies, single-chain antibodies, and antibody fragments (e.g., Fab, F(ab')₂, and Fv). Also, the antibody is any class of immunoglobulin molecule, for example, IgG, IgE, IgM, IgA, IgD, or 65 IgY, or any subclass, for example, IgG1, IgG2, IgG3, IgG4, IgA1, or IgA2.

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The antibody may be further modified by acetylation, formylation, amidation, phosphorylation, PEGylation, or the like, in addition to glycosylation.

Hereinafter, preparation examples of various antibodies will be shown.

When the antibody of the present invention is a monoclonal antibody, for example, a breast cancer cell line SK-BR-3 expressing CAPRIN-1 is administered to each mouse for immunization. The spleen is extracted from this mouse. After separation of spleen cells, the cells are fused with mouse myeloma cells. Clones producing antibodies having a cancer cell growth inhibitory effect are selected from among the obtained fusion cells (hybridomas). Alternatively, clones producing antibodies binding to a polypeptide consisting of the amino acid sequence set forth in SEQ ID NO: 5 or a polypeptide consisting of an amino acid sequence having 80% or higher sequence identity to the amino acid sequence may be selected. The hybridomas producing monoclonal antibodies having a cancer cell growth inhibitory effect or the hybridomas producing monoclonal antibodies against the polypeptide of SEQ ID NO: 5 or the like are isolated and cultured. The antibody of the present invention can be prepared by purification from the culture supernatant according to a general affinity purification method.

The monoclonal antibody-producing hybridomas may be prepared, for example, as follows: first, animals are immunized with sensitizing antigens according to a method known in the art. This immunization method generally involves intraperitoneally or subcutaneously injecting the sensitizing antigens to mammals. Specifically, the sensitizing antigens are diluted with or suspended in PBS (phosphate-buffered saline), physiological saline, or the like into an appropriate amount and then mixed, if desired, with an appropriate amount of a conventional adjuvant, for example, a complete Freund's adjuvant. After emulsification, the resulting emulsion is administered to each mammal several times every 4 to 21 days. Alternatively, an appropriate carrier may be used for the immunization with sensitizing antigens.

After confirmation of a rise in the level of the desired antibody in the serum of the mammal thus immunized, immunocytes are collected from the mammal and subjected to cell fusion. Preferred examples of the immunocytes particularly include spleen cells.

Mammalian myeloma cells are used as partner parent cells to be fused with the immunocytes. Various cell lines known in the art, for example, P3U1 (P3-X63Ag8U1), P3 (P3x63Ag8.653) (J. Immunol. (1979) 123, 1548-1550), P3x63Ag8U.1 (Current Topics in Microbiology and Immunology (1978) 81, 1-7), NS-1 (Kohler. G. and Milstein, C. Eur. J. Immunol. (1976) 6, 511-519), MPC-11 (Margulies. D. H. et al., Cell (1976) 8, 405-415), SP2/0 (Shulman, M. et al., Nature (1978) 276, 269-270), FO (deSt. Groth, S. F. et al., J. Immunol. Methods (1980) 35, 1-21), 5194 (Trowbridge, I. S. J. Exp. Med. (1978) 148, 313-323), and R210 (Galfre, G. et al., Nature (1979) 277, 131-133), are preferably used as the myeloma cells.

The cell fusion between the immunocytes and the myeloma cells can be performed basically according to a method known in the art, for example, the method of Kohler and Milstein (Kohler, G. and Milstein, C. Methods Enzymol. (1981) 73, 3-46).

More specifically, the cell fusion is carried out, for example, in the presence of a cell fusion promoter in a conventional nutrient medium. For example, polyethylene glycol (PEG) or hemagglutinating virus of Japan (HVJ) is used as

the fusion promoter. If desired, an auxiliary such as dimethyl sulfoxide may be further added for use in order to enhance fusion efficiency.

The ratio between the immunocytes and the myeloma cells used can be arbitrarily set. For example, the amount of the 5 immunocytes is preferably set to 1 to 10 times the amount of the myeloma cells. Examples of the medium that can be used in the cell fusion include RPMI1640 and MEM media suitable for the growth of the myeloma cell lines as well as conventional media for use in this type of cell culture. In 10 addition, a serum supplement such as fetal calf serum (FCS) may be used in combination with these media.

For the cell fusion, the immunocytes and the myeloma cells are well mixed in a predetermined amount of the medium. A PEG solution (average molecular weight: for example, 15 approximately 1000 to 6000) preheated to approximately 37° C. is usually added to the mixture at a concentration of 30 to 60% (w/v) and mixed therewith to form the hybridomas of interest. Subsequently, procedures of sequentially adding an appropriate medium and removing the supernatant by centrifugation are preferably repeated to remove cell fusion agents or the like unfavorable for the growth of the hybridomas.

The hybridomas thus obtained are cultured in a conventional selective medium, for example, a HAT medium (medium containing hypoxanthine, aminopterin, and thymidine) for selection. This culture in the HAT medium is continued for a period (usually, several days to several weeks) sufficient for the death of cells (non-fused cells) other than the hybridomas of interest. Subsequently, hybridomas producing the antibody of interest are screened for and cloned as single clones by a conventional limiting dilution method.

In addition to such obtainment of the hybridomas by the immunization of non-human animals with antigens, hybridomas producing human antibodies having the desired activity 35 (e.g., cell growth inhibitory activity) may be obtained by sensitizing human lymphocytes, for example, EB virus-infected human lymphocytes, with proteins, protein-expressing cells, or lysates thereof in vitro and fusing the sensitized lymphocytes with human-derived myeloma cells capable of 40 dividing permanently, for example, U266 (Registration No. TIB196).

The monoclonal antibody-producing hybridomas thus prepared can be subcultured in a conventional medium and can also be stored for a long period in liquid nitrogen.

Specifically, the desired antigens or cells expressing the desired antigens are used as sensitizing antigens in immunization according to a conventional immunization method. The obtained immunocytes are fused with parent cells known in the art according to a conventional cell fusion method. 50 Monoclonal antibody-producing cells (hybridomas) can be screened for by a conventional screening method to prepare the antibody of interest.

Another example of the antibody that may be used in the present invention is a polyclonal antibody. The polyclonal 55 antibody can be obtained, for example, as follows:

Serum is obtained from small animals such as mice, human antibody-producing mice, or rabbits immunized with natural CAPRIN-1 proteins or recombinant CAPRIN-1 proteins expressed as fusion proteins with GST or the like in microorganisms such as *E. coli*, or partial peptides thereof. Alternatively, serum may be obtained from mammals immunized with CAPRIN-1 fragments serving as sensitizing antigens, i.e., a polypeptide comprising the amino acid sequence set forth in SEQ ID NO: 5 or an amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to

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the amino acid sequence (preferably, a polypeptide consisting of any of these amino acid sequences), or a polypeptide comprising an epitope (preferably, consisting of the epitope) consisting of approximately 7 to 12 consecutive amino acids, for example, 8 to 11 consecutive amino acids, in the amino acid sequence set forth in SEQ ID NO: 5 or the amino acid sequence having 80% or higher, preferably 85% or higher, more preferably 90% or higher, further preferably 95% or higher sequence identity to the amino acid sequence. The serum thus obtained can be purified using, for example, ammonium sulfate precipitation, protein A or protein G columns, DEAE ion-exchange chromatography, or affinity columns coupled with CAPRIN-1 proteins or synthetic peptides to prepare anti-CAPRIN-1 polyclonal antibodies. The polyclonal antibody of the present invention includes antibodies obtained from human antibody-producing animals (e.g., mice) immunized with CAPRIN-1 proteins.

In this context, for example, KM mice (Kirin Pharma Co., Ltd./Medarex) and Xeno mice (Amgen Inc.) are known as the human antibody-producing mice (e.g., International Publication Nos. WO02/43478 and WO02/092812). Complete human polyclonal antibodies can be obtained from the blood of such mice immunized with CAPRIN-1 proteins or fragments thereof. Alternatively, spleen cells may be isolated from the mice thus immunized and fused with myeloma cells. In this way, human monoclonal antibodies can be obtained.

The antigens can be prepared according to, for example, a method using animal cells (JP Patent Publication (Kohyo) No. 2007-530068 A) or a method using baculovirus (e.g., International Publication No. WO98/46777). Antigens having low immunogenicity can be bound to immunogenic macromolecules such as albumin for immunization. The antigens may be administered together with adjuvants for immunization.

Alternatively, the antibody of the present invention may be obtained as a genetically recombinant antibody that is produced using a gene recombination technique which involves: cloning a gene of the antibody from hybridomas; incorporating the antibody gene into appropriate vectors; and introducing the vectors into hosts (see, e.g., Carl, A. K. Borrebaeck, James, W. Larrick, THERAPEUTIC MONOCLONAL ANTIBODIES, Published in the United Kingdom by MAC-MILLAN PUBLISHERS LTD, 1990). Specifically, antibody variable region (V region) cDNAs are synthesized from the mRNAs of hybridomas using reverse transcriptase. After obtainment of DNAs encoding the antibody V regions of interest, the DNAs are ligated with DNAs encoding the desired antibody constant regions (C regions). The resulting ligation products are incorporated into expression vectors. Alternatively, the antibody V region-encoding DNAs may be incorporated into expression vectors containing antibody C region DNAs. These DNAs are incorporated into the expression vectors so as to be expressed under the control of expression control regions, for example, an enhancer and a promoter. Next, host cells can be transformed with the resulting expression vectors and allowed to express antibodies.

The anti-CAPRIN-1 antibody of the present invention is preferably a monoclonal antibody. Alternatively, the anti-CAPRIN-1 antibody of the present invention may be a polyclonal antibody, a genetically engineered antibody (chimeric antibody, humanized antibody, etc.), or the like.

The monoclonal antibody includes human monoclonal antibodies, non-human animal monoclonal antibodies (e.g., mouse, rat, rabbit, and chicken monoclonal antibodies), chimeric monoclonal antibodies, and the like. The monoclonal antibody may be prepared by the culture of hybridomas obtained by the fusion between spleen cells from non-human

mammals (e.g., mice, human antibody-producing mice, chickens, or rabbits) immunized with CAPRIN-1 proteins or fragments thereof and myeloma cells. The chimeric antibody is an antibody prepared from a combination of sequences derived from different animals and is, for example, an antibody composed of mouse antibody heavy and light chain variable regions and human antibody heavy and light chain constant regions. The chimeric antibody can be prepared using a method known in the art which involves, for example: ligating DNAs encoding mouse antibody V regions with 10 DNAs encoding human antibody C regions; incorporating the resulting ligation products into expression vectors; and introducing the vectors into hosts so that antibodies are produced.

Monoclonal antibodies that have immunological reactivity with the partial CAPRIN-1 polypeptide consisting of the 15 amino acid sequence set forth in SEQ ID NO: 5 and have an antitumor effect are prepared by a method described later in Examples.

The humanized antibody, also called reshaped human antibody, is an engineered antibody. The humanized antibody is 20 constructed by introducing antibody CDRs derived from an immunized animal into human antibody complementarity determining regions. A general gene recombination approach therefor is also known.

Specifically, DNA sequences designed so as to link, for 25 example, mouse, rabbit, or chicken antibody CDRs, and human antibody framework regions (FRs) are synthesized by PCR using several prepared oligonucleotides having terminal portions overlapping with each other. The obtained DNAs are ligated with DNAs encoding human antibody constant 30 regions. Subsequently, the resulting ligation products are incorporated into expression vectors, which are then introduced into hosts for antibody production to obtain the antibody of interest (see European Patent Application Publication No. EP239400 and International Publication No. WO96/ 35 02576). The human antibody FRs connected via CDRs are selected such that the complementarity determining regions form a favorable antigen-binding site. If necessary, amino acids in the framework regions of antibody variable regions may be substituted such that the complementarity determin- 40 ing regions of the resulting reshaped human antibody form an appropriate antigen-binding site (Sato K. et al., Cancer Research 1993, 53: 851-856). In addition, these FRs may be replaced with framework regions derived from various human antibodies of class or subclass different therefrom (see 45 International Publication No. WO99/51743).

The human antibody framework regions connected via CDRs are selected such that the complementarity determining regions form a favorable antigen-binding site. If necessary, amino acids in the framework regions of antibody variable regions may be substituted such that the complementarity determining regions of the resulting reshaped human antibody form an appropriate antigen-binding site (Sato K. et al., Cancer Research 1993, 53: 851-856).

Amino acids in variable regions (e.g., FRs) or constant 55 regions of the chimeric antibody or the humanized antibody thus prepared may be substituted, for example, by other amino acids.

The amino acid substitution is the substitution of, for example, less than 15, less than 10, 8 or less, 7 or less, 6 or 60 less, 5 or less, 4 or less, 3 or less, or 2 or less amino acids, preferably 1 to 5 amino acids, more preferably 1 or 2 amino acids. The substituted antibody should be functionally equivalent to an unsubstituted antibody. The substitution is desirably conservative amino acid substitution, which is the 65 substitution between amino acids similar in properties such as charge, side chains, polarity, and aromaticity. The amino

acids can be classified in terms of similar properties into, for example: basic amino acids (arginine, lysine, and histidine); acidic amino acids (aspartic acid and glutamic acid); uncharged polar amino acids (glycine, asparagine, glutamine, serine, threonine, cysteine, and tyrosine); nonpolar amino acids (leucine, isoleucine, alanine, valine, proline, phenylalanine, tryptophan, and methionine); branched amino acids (leucine, valine, and isoleucine); and aromatic amino acids (phenylalanine, tyrosine, tryptophan, and histidine).

Examples of modified antibodies can include antibodies bound with various molecules such as polyethylene glycol (PEG). In the modified antibody of the present invention, the substance to be bound is not limited. In order to obtain such a modified antibody, the obtained antibody can be chemically modified. A method therefor has already been established in the art.

In this context, the phrase "functionally equivalent" means that an antibody concerned has biological or biochemical activity similar to that of the antibody of the present invention, specifically, the antibody concerned has the function of damaging tumor and essentially causes no rejection when applied to humans, for example. Examples of such activity can include cell growth inhibitory activity and binding activity.

A method for preparing a polypeptide functionally equivalent to a certain polypeptide, which involves introducing a mutation into a polypeptide, is well known to those skilled in the art. For example, those skilled in the art can appropriately introduce a mutation into the antibody of the present invention using site-directed mutagenesis (Hashimoto-Gotoh, T. et al., (1995) Gene 152, 271-275; Zoller, M J., and Smith, M. (1983) Methods Enzymol. 100, 468-500; Kramer, W. et al., (1984) Nucleic Acids Res. 12, 9441-9456; Kramer, W. and Fritz, H J., (1987) Methods Enzymol. 154, 350-367; Kunkel, T A., (1985) Proc. Natl. Acad. Sci. USA. 82, 488-492; and Kunkel (1988) Methods Enzymol. 85, 2763-2766) or the like, thereby preparing an antibody functionally equivalent to the antibody of the present invention.

The antibody that recognizes an epitope of a CAPRIN-1 protein or a CAPRIN-1 fragment polypeptide comprising the epitope can be obtained by a method generally known to those skilled in the art. For example, the antibody can be obtained by a method which involves determining the epitope of the CAPRIN-1 protein recognized by the obtained anti-CA-PRIN-1 antibody having a cancer cell growth inhibitory effect by a conventional method (e.g., epitope mapping or an epitope identification method described later) and preparing an antibody using a polypeptide having an amino acid sequence contained in the epitope as an immunogen, or a method which involves determining an epitope for an antibody prepared by a conventional method and selecting an antibody that recognizes the same epitope as that for the anti-CAPRIN-1 antibody. In this context, the "epitope" refers to a polypeptide fragment having antigenicity or immunogenicity in mammals, preferably humans. Its smallest unit consists of approximately 7 to 12 amino acids, preferably 8 to 11 amino acids.

The antibody of the present invention is an antibody having immunological reactivity with CAPRIN-1, an antibody that specifically recognizes CAPRIN-1, or an antibody that specifically binds to CAPRIN-1 and exhibits cytotoxic activity against cancer or a tumor growth inhibitory effect. The antibody is preferably an antibody having a structure that causes little or no rejection in recipient animals. Examples of such antibodies include human antibodies, humanized antibodies, chimeric antibodies (e.g., human-mouse chimeric antibodies), single-chain antibodies, and bispecific antibodies when the recipient animals are humans. These antibodies have

heavy and light chain variable regions derived from a human antibody or have heavy and light chain variable regions with complementarity determining regions (CDR1, CDR2, and CDR3) derived from a non-human animal antibody and framework regions (FR1, FR2, FR3, and FR4) derived from a 5 human antibody. Alternatively, these antibodies are recombinant antibodies having heavy and light chain variable regions derived from a non-human animal antibody and heavy and light chain constant regions derived from a human antibody. The antibody of the present invention is preferably the former 10 two antibodies.

Such recombinant antibodies can be prepared as follows: DNAs encoding monoclonal antibodies (e.g., human, mouse, rat, rabbit, and chicken monoclonal antibodies) against human CAPRIN-1 are cloned from antibody-producing cells 15 such as hybridomas and used as templates in RT-PCR or the like to prepare DNAs encoding the light and heavy chain variable regions of the antibodies. The respective sequences of the light and heavy chain variable regions, the respective sequences of CDR1, CDR2, and CDR3 in each region, or the 20 respective sequences of FR1, FR2, FR3, and FR4 in each region can be determined on the basis of, for example, the Kabat EU numbering system (Kabat et al., Sequences of Proteins of Immunological Interest, 5th Ed. Public Health Service, National Institute of Health, Bethesda, Md. (1991)). 25

Such a DNA encoding each variable region or a DNA encoding each CDR is prepared using a gene recombination technique (Sambrook et al., Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory Press (1989)) or a DNA synthesizer. In this context, the human monoclonal 30 antibody-producing hybridomas can be prepared by immunizing human antibody-producing animals (e.g., mice) with human CAPRIN-1 and then fusing spleen cells excised from the immunized animals with myeloma cells. Aside from this, DNAs encoding human antibody-derived light or heavy chain variable and constant regions are prepared, if necessary, using a gene recombination technique or a DNA synthesizer.

For the humanized antibody, DNAs in which the CDR coding sequences in DNAs encoding human antibody-derived light or heavy chain variable regions are substituted by 40 corresponding CDR coding sequences of a non-human animal (e.g., mouse, rat, rabbit, or chicken)-derived antibody can be prepared and ligated with the DNAs encoding human antibody-derived light or heavy chain constant regions to prepare a DNA encoding the humanized antibody.

For the chimeric antibody, DNAs encoding light or heavy chain variable regions of a non-human animal (e.g., mouse, rat, rabbit, or chicken)-derived antibody can be ligated with DNAs encoding human antibody-derived light or heavy chain constant regions to prepare a DNA encoding the chimeric 50 antibody.

The single-chain antibody refers to an antibody comprising heavy and light chain variable regions linearly linked to each other via a linker. A DNA encoding the single-chain antibody can be prepared by ligating a DNA encoding the heavy chain 55 variable region, a DNA encoding the linker, and a DNA encoding the light chain variable region. In this context, the heavy and light chain variable regions are both derived from a human antibody or derived from a human antibody or derived from a human antibody having CDRs alone substituted by CDRs of a non-human animal (e.g., mouse, rat, rabbit, or chicken)-derived antibody. The linker consists of 12 to 19 amino acids. Examples thereof include $(G_4S)_3$ consisting of 15 amino acids (G. B. Kim et al., Protein Engineering Design and Selection 2007, 20 (9): 425-432)

The bispecific antibody (e.g., diabody) refers to an antibody capable of specifically binding to two different epitopes. 14

A DNA encoding the bispecific antibody can be prepared by ligating, for example, a DNA encoding a heavy chain variable region A, a DNA encoding a light chain variable region B, a DNA encoding a heavy chain variable region B, and a DNA encoding a light chain variable region A in this order (provided that the DNA encoding a light chain variable region B and the DNA encoding a heavy chain variable region B are ligated via a DNA encoding a linker as described above). In this context, the heavy and light chain variable regions are all derived from a human antibody or derived from a human antibody having CDRs alone substituted by CDRs of a non-human animal (e.g., mouse, rat, rabbit; or chicken)-derived antibody.

The recombinant DNAs thus prepared can be incorporated into one or more appropriate vectors, which are then introduced into host cells (e.g., mammalian cells, yeast cells, and insect cells) so that the DNAs are (co)expressed to produce recombinant antibodies (P. J. Delves., ANTIBODY PRODUCTION ESSENTIAL TECHNIQUES., 1997 WILEY, P. Shepherd and C. Dean., Monoclonal Antibodies., 2000 OXFORD UNIVERSITY PRESS; and J. W. Goding., Monoclonal Antibodies: principles and practice., 1993 ACA-DEMIC PRESS).

Proteins of Immunological Interest, 5th Ed. Public Health
Service, National Institute of Health, Bethesda, Md. (1991)).

Such a DNA encoding each variable region or a DNA

Examples of the antibody of the present invention prepared by any of the methods described above include the following antibodies (a) and (b):

(a) an antibody comprising a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 8, 9, and 10 and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (e.g., an antibody constituted by a heavy chain variable region of SEQ ID NO: 11 and a light chain variable region of SEQ ID NO: 15); and

(b) an antibody comprising a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 18, 19, and 20 and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 22, 23, and 24 (e.g., an antibody constituted by a heavy chain variable region of SEQ ID NO: 21 and a light chain variable region of SEQ ID NO: 25).

In this context, the amino acid sequences represented by SEQ ID NOs: 8, 9, and 10 correspond to heavy chain variable region CDR1, CDR2, and CDR3, respectively. The amino acid sequences represented by SEQ ID NOs: 12, 13, and 14 correspond to light chain variable region CDR1, CDR2, and CDR3, respectively. The amino acid sequences represented by SEQ ID NOs: 18, 19, and 20 correspond to heavy chain variable region CDR1, CDR2, and CDR3, respectively. The amino acid sequences represented by SEQ ID NOs: 22, 23, and 24 correspond to light chain variable region CDR1, CDR2, and CDR3, respectively.

Examples of the humanized antibody, the chimeric antibody, the single-chain antibody, or the bispecific antibody of the present invention include the following antibodies (i) to (iii), for example, in the form of (a):

(i) an antibody comprising a heavy chain variable region comprising CDR1, CDR2, and CDR3 consisting of the amino acid sequences of SEQ ID NOs: 8, 9, and 10, respectively, and humanized antibody-derived framework regions and a light chain variable region comprising CDR1, CDR2, and CDR3 consisting of the amino acid sequences of SEQ ID NOs: 12, 13, and 14, respectively, and human antibody-derived framework regions;

(ii) an antibody comprising a heavy chain comprising a heavy chain variable region comprising CDR1, CDR2, and CDR3 consisting of the amino acid sequences of SEQ ID NOs: 8, 9, and 10, respectively, and human antibody-derived

framework regions, and a human antibody-derived heavy chain constant region, and a light chain comprising a light chain variable region comprising CDR1, CDR2, and CDR3 consisting of the amino acid sequences of SEQ ID NOs: 12, 13, and 14, respectively, and human antibody-derived framework regions, and a human antibody-derived a light chain constant region; and

(iii) an antibody comprising a heavy chain comprising a heavy chain variable region comprising the amino acid sequence of SEQ ID NO: 11 and a human antibody-derived heavy chain constant region, and a light chain comprising a light chain variable region comprising the amino acid sequence of SEQ ID NO: 15 and a human antibody-derived light chain constant region.

The sequences of the constant and variable regions of 15 human antibody heavy and light chains are available from, for example, NCBI (USA; GenBank, UniGene, etc.). For example, the following sequences can be referred to: Registration No. J00228 for a human IgG1 heavy chain constant region; Registration No. J00230 for a human IgG2 heavy chain constant region; Registration No. X03604 for a human IgG3 heavy chain constant region; Registration No. K01316 for a human IgG4 heavy chain constant region; Registration Nos. V00557, X64135, X64133, etc. for a human light chain κ constant region; and Registration Nos. X64132, X64134, 25 etc. for a human light chain λ constant region.

Preferably, these antibodies have cytotoxic activity and can thereby exert an antitumor. effect.

The above particular sequences of the heavy and light chain variable regions and CDRs in each antibody are provided merely for illustrative purposes. It is obvious that the antibody of the present invention is not limited by the particular sequences. Hybridomas capable of producing antihuman CAPRIN-1 human antibodies or non-human animal antibodies (e.g., mouse antibodies) different from those 35 described above are prepared, and monoclonal antibodies produced by the hybridomas are recovered and assessed as being (or being not) the antibodies of interest with immunological binding activity against human CAPRIN-1 and cytotoxic activity as indexes. The monoclonal antibody-produc- 40 ing hybridomas of interest are thereby identified. Then, DNAs encoding the heavy and light chain variable regions of the antibodies of interest are produced from the hybridomas and sequenced, as described above. The DNAs are used for the preparation of the different antibodies.

The above antibodies may each have the substitution, deletion, or addition of one or several amino acids, particularly in a framework region sequence and/or a constant region sequence, as long as the antibody has such specificity that it can specifically recognize CAPRIN-1. In this context, the 50 term "several" means preferably 2 to 5, more preferably 2 or 3.

The affinity constant Ka (k_{om}/k_{off}) of the antibody of the present invention for a CAPRIN-1 protein or a fragment thereof is preferably at least $10^7 \, \text{M}^{-1}$, at least $10^8 \, \text{M}^{-1}$, at least $10^8 \, \text{M}^{-1}$, at least $10^9 \, \text{M}^{-1}$, at least $10^{10} \, \text{M}^{-1}$, at least $10^{10} \, \text{M}^{-1}$, at least $10^{11} \, \text{M}^{-1}$, at least $10^{12} \, \text{M}^{-1}$, at least $10^{12} \, \text{M}^{-1}$, at least $10^{12} \, \text{M}^{-1}$, or at least $10^{13} \, \text{M}^{-1}$.

The antibody of the present invention can be conjugated with an antitumor agent. The conjugation of the antibody with 60 the antitumor agent can be performed via a spacer having a group (e.g., a succinimidyl group, a formyl group, a 2-pyridyldithio group, a maleimidyl group, an alkoxycarbonyl group, or a hydroxy group) reactive with an amino group, a carboxyl group, a hydroxy group, a thiol group, or the like. 65

Examples of the antitumor agent include the following antitumor agents publicly known in literatures, etc.: pacli-

taxel, doxorubicin, daunorubicin, cyclophosphamide, methotrexate, 5-fluorouracil, thiotepa, busulfan; improsulfan, piposulfan, benzodopa, carboquone, meturedopa, uredopa, altretamine, triethylenemelamine, triethylenephosphoramide, triethylenethiophosphoramide, trimethylolomelamine, bullatacin, bullatacinone, camptothecin, bryostatin, callystatin, cryptophycin 1, cryptophycin 8, dolastatin, duocarmycin, eleutherobin, pancratistatin, sarcodictyin, spongistatin, chlorambucil, chlomaphazine, cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard, carmustine, chlorozotocin, fotemustine, lomustine, nimustine, ranimustine, calicheamicin, dynemicin, clodronate, esperamicin, aclacinomycin, actinomycin, authramycin, azaserine, bleomycin, cactinomycin, carabicin, carminomycin, carzinophilin, chromomycin, dactinomycin, detorbicin, 6-diazo-5-oxo-L-norleucine, Adriamycin, epirubicin, esorubicin, idarubicin, marcellomycin, mitomycin C, mycophenolic acid, nogalamycin, olivomycin, peplomycin, potfiromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin, denopterin, pteropterin, trimetrexate, fludarabine, 6-mercaptopurine, thiamiprine, thioguanine, ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine, androgens (e.g., calusterone, dromostanolone propionate, epitiostanol, mepitiostane, and testolactone), aminoglutethimide, mitotane, trilostane, frolinic acid, aceglatone, aldophosphamide glycoside, aminolevulinic acid, eniluracil, amsacrine, bestrabucil, bisantrene, edatraxate, defofamine, demecolcine, diaziquone, elfornithine, elliptinium acetate, epothilone, etoglucid, lentinan, lonidamine, maytansine, ansamitocin, mitoguazone, mitoxantrone, mopidanmol, nitraerine, pentostatin, phenamet, pirarubicin, losoxantrone, podophyllinic acid, 2-ethylhydrazide, procarbazine, razoxane, rhizoxin, schizophyllan, spirogermanium, tenuazonic acid, triaziquone, roridin A, anguidine, urethane, vindesine, dacarbazine, mannomustine, mitobronitol, mitolactol, pipobroman, gacytosine, docetaxel, chlorambucil, gemcitabine, 6-thioguanine, mercaptopurine, cisplatin, oxaliplatin, carboplatin, vinblastine, etoposide, ifosfamide, mitoxantrone, vincristine, vinorelbine, novantrone, teniposide, edatrexate, daunomycin, aminopterin, Xeloda, ibandronate, irinotecan, topoisomerase inhibitors, difluoromethylornithine (DMFO), retinoic acid, capecitabine, and pharmaceutically acceptable salts and derivatives thereof.

Alternatively, the antibody of the present invention can be administered in combination with an antitumor agent to produce a higher therapeutic effect. This approach is adaptable to a patient with cancer expressing CAPRIN-1 either before or after surgical operation. This approach can be applied, particularly after surgery, to CAPRIN-1-expressing cancer, which has been treated conventionally with an antitumor agent alone, to produce higher prevention of cancer recurrence or prolongation of survival time.

Examples of the antitumor agent used in the combined administration with the antibody of the present invention also include the following antitumor agents publicly known in literatures, etc.: paclitaxel, doxorubicin, daunorubicin, cyclophosphamide, methotrexate, 5-fluorouracil, thiotepa, busulfan, improsulfan, piposulfan, benzodopa, carboquone, meturedopa, uredopa, altretamine, triethylenemelamine, triethylenephosphoramide, triethylenethiophosphoramide, triemethylolomelamine, bullatacin, bullatacinone, camptothecin, bryostatin, callystatin, cryptophycin 1, cryptophycin 8, dolastatin, duocarmycin, eleutherobin, pancratistatin, sarcodictyin, spongistatin, chlorambucil, chlornaphazine,

cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard, carmustine, chlorozotocin, fotemustine, lomustine, nimustine, ranimustine, calicheamicin, dynemi- 5 cin, clodronate, esperamicin, aclacinomycin, actinomycin, authramycin, azaserine, bleomycin, cactinomycin, carabicin, carminomycin, carzinophilin, chromomycin, dactinomycin, detorbicin, 6-diazo-5-oxo-L-norleucine, Adriamycin, epirubicin, esorubicin, idarubicin, marcellomycin, mitomycin C, mycophenolic acid, nogalamycin, olivomycin, peplomycin, potfiromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin, denopterin, pteropterin, trimetrexate, fludarabine, 6-mercaptopurine, thiamiprine, thioguanine, ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine, calusterone, dromostanolone propionate, epitiostanol, mepitiostane, testolactone, aminoglutethimide, mitotane, trilostane, frolinic acid, aceglatone, aldophosphamide glycoside, aminolevulinic 20 acid, eniluracil, amsacrine, bestrabucil, bisantrene, edatraxate, defofamine, demecolcine, diaziquone, elfornithine, elliptinium acetate, epothilone, etoglucid, lentinan, lonidamine, maytansine, ansamitocin, mitoguazone, mitoxantrone, mopidanmol, nitraerine, pentostatin, phenamet, pirarubicin, losox-25 antrone, podophyllinic acid, 2-ethylhydrazide, procarbazine, razoxane, rhizoxin, schizophyllan, spirogermanium, tenuazonic acid, triaziquone, roridin A, anguidine, urethane, vindesine, dacarbazine, mannomustine, mitobronitol, mitolactol, pipobroman, gacytosine, docetaxel, chlorambucil, 30 gemcitabine, 6-thioguanine, mercaptopurine, cisplatin, oxaliplatin, carboplatin, vinblastine, etoposide, ifosfamide, mitoxantrone, vincristine, vinorelbine, novantrone, teniposide, edatrexate, daunomycin, aminopterin, Xeloda, ibandronate, irinotecan, topoisomerase inhibitors, difluoromethy- 35 lornithine (DMFO), retinoic acid, capecitabine, and pharmaceutically acceptable salts (known in the art) and derivatives (known in the art) thereof. Of these antitumor agents, cyclophosphamide, paclitaxel, docetaxel, or vinorelbine is particularly preferably used.

Alternatively, the antibody of the present invention may be bound to a radioisotope publicly known in literatures, etc., such as, ²¹¹At, ¹³¹I, ¹²⁵I, ⁹⁰Y, ¹⁸⁶Re, ¹⁸⁸Re, ¹⁵³SM, ²¹²Bi, ³²P, ¹⁷⁵Lu, ¹⁷⁶Lu, ⁸⁹Sr, ⁶⁴Cu, or ¹¹¹In (Hideo Saji, YAKUGAKU ZASSHI 128 (3) 323-332, 8 (2008), Jpn). A radioisotope 45 effective for the treatment or diagnosis of tumor is desirable. Such a radioisotope is also included in the antitumor agent according to the present invention.

<Identification of Epitope>

As shown in Examples below, the antibody of the present 50 invention binds to an epitope in the amino acid sequence set forth in SEQ ID NO: 5. One example of a method for confirming an epitope for the antibody of the present invention includes a method which involves immobilizing an epitope in the polypeptide of SEQ ID NO: 5 onto a plate and evaluating 55 the antibody for its reactivity with this epitope. Specifically, an epitope in the polypeptide of SEQ ID NO: 5 is immobilized through reaction onto a plate attached with electron-withdrawing functional groups via spacers such as oligoethylene glycol. The antibody of the present invention can be reacted 60 with the plate and evaluated for its reactivity with the epitope through reaction with a labeled (e.g., horseradish peroxidase (HRP)-labeled) secondary antibody binding to the antibody of the present invention, i.e., the epitope to which the antibody of the present invention binds can be confirmed. The epitope 65 in the polypeptide of SEQ ID NO: 5 used in the immobilization onto a plate is a sequence itself comprising at least the

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epitope in the sequence of SEQ ID NO: 5 or a modified portion thereof (e.g., N-terminal or C-terminal residues modified with several arbitrary amino acids or a protein such as KLH or a (poly)peptide modified with a MAP protein). The antibody of the present invention needs only to bind to any of these (poly)peptides.

On the other hand, even the antibody of the present invention may be unreactive with the polypeptide of SEQ ID NO: 5, i.e., the epitope may not be confirmed, in the above method. In this case, the antibody is reacted with an antigen under solution conditions that facilitate the binding between the antigen and the antibody. After obtainment of an antigenantibody complex by an immunoprecipitation method, a partial polypeptide bound with the antibody can be separated and examined for its amino acid sequence to confirm the epitope for the antibody of the present invention. In this context, the antigen is, for example, the polypeptide of SEQ ID NO: 5 itself or a modified portion thereof. Alternatively, even a CAPRIN-1 protein may be used as long as the epitope reactive with the antibody of the present invention can be confirmed by the above method.

<Antitumor Effect>

The antitumor effect of the anti-CAPRIN-1 antibody used in the present invention on CAPRIN-1-expressing cancer cells seems to be brought about by the following mechanism: effector cell-mediated antibody-dependent cellular. cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) against the CAPRIN-1-expressing cells. However, this mechanism is not intended to limit the scope of the present invention.

The antitumor effect based on the mechanism is known to correlate with the number of target molecules expressed on the surface of cancer cells to which the antibody binds (Niwa R., Clinical Cancer Research 2005 Mar. 15; 11 (6): 2327-2336). The number of target molecules expressed on the surface of cancer cells can be examined using an existing assay kit capable of measuring the number of cell surface molecules. Specifically, the number of target molecules to which the antibody binds can be determined by: reacting primary antibodies such as antibodies against the target molecules with cancer cells; reacting therewith fluorescently labeled antibodies against the primary antibodies together with beads for a calibration curve with the known number of molecules; and measuring the mean fluorescence intensity of the samples to obtain a calibration curve.

Thus, the anti-CAPRIN-1 antibody used in the present invention can be evaluated for its activity, as specifically shown in Examples below, by assaying the ADCC or CDC activity against CAPRIN-1-expressing cancer cells ex vivo or by examining the number of CAPRIN-1 molecules expressed on the surface of cancer cells using the anti-CAPRIN-1 anti-body according to the present invention as a primary anti-body.

The anti-CAPRIN-1 antibody used in the present invention binds to a CAPRIN-1 protein on cancer cells and exhibits an antitumor effect through the activity. Thus, the anti-CA-PRIN-1 antibody of the present invention is presumably useful in the treatment or prevention of cancer. Specifically, the present invention provides a pharmaceutical composition for treatment and/or prevention of cancer, comprising the anti-CAPRIN-1 antibody as an active ingredient. The anti-CA-PRIN-1 antibody used for the purpose of administration to human bodies (antibody therapy) is preferably a human antibody or a humanized antibody for reducing immunogenicity.

An anti-CAPRIN-1 antibody with higher binding affinity for a CAPRIN-1 protein on cancer cell surface exerts stronger antitumor activity. Thus, the antibody of the present invention

has high binding affinity for the CAPRIN-1 protein and can therefore be expected to have a stronger antitumor effect. Accordingly, the antibody of the present invention is adaptable to a pharmaceutical composition intended for the treatment and/or prevention of cancer. Such high binding affinity of the antibody of the present invention is preferably at least $10^7\,M^{-1}$, at least $10^8\,M^{-1}$, at least $5\times10^8\,M^{-1}$, at least $10^9\,M^{-1}$, at least $5\times10^9\,M^{-1}$, at least $10^{10}\,M^{-1}$, at least $5\times10^{10}\,M^{-1}$, at least $10^{11}\,M^{-1}$, at least $10^{12}\,M^{-1}$, or at least $10^{13}\,M^{-1}$, in terms of an association constant (affinity 10 constant) Ka (k_{on}/k_{off}) , as described above.

The anti-CAPRIN-1 antibody binding to a larger number of CAPRIN-1 molecules on cancer cell surface produces stronger antitumor activity. Desirably, the number of CAPRIN-1 molecules in assay using the anti-CAPRIN-1 15 antibody of the present invention is 10⁴ or more, preferably 10⁵ or more, per cancer cell to which the antibody binds in expectation of the antitumor effect. Tumor (cancer cells) having a large number of CAPRIN-1 molecules on the cell surface is particularly preferred as cancer to receive the antibody 20 of the present invention.

<Binding to Antigen-expressing Cell>

The ability of the antibody to bind to CAPRIN-1 can be determined by use of binding assay using, for example, ELISA, Western blot, immunofluorescence, and flow cytom- 25 etry analysis, as described in Examples.

<Immunohistochemical Staining>

The antibody that recognizes CAPRIN-1 can be tested for its reactivity with CAPRIN-1 by an immunohistochemical method well known to those skilled in the art using a 30 paraformaldehyde- or acetone-fixed frozen section or paraformaldehyde-fixed paraffin-embedded section of a tissue obtained from a patient during surgical operation or from an animal carrying a xenograft tissue inoculated with a cell line expressing CAPRIN-1 either spontaneously or after 35 transfection.

For immunohistochemical staining, the antibody reactive with CAPRIN-1 can be stained by various methods. For example, the antibody can be visualized through reaction with a horseradish peroxidase-conjugated goat anti-mouse 40 antibody, goat anti-rabbit antibody, or goat anti-chicken antibody.

<Pharmaceutical Composition and Method for Treating and/ or Preventing Cancer>

A target of the pharmaceutical composition for treatment 45 and/or prevention of cancer of the present invention is not particularly limited as long as the target is cancer (cells) expressing a CAPRIN-1 gene.

The terms "tumor" and "cancer" used herein mean malignant neoplasm and are used interchangeably with each other. 50

The cancer targeted in the present invention is cancer expressing a gene encoding a CAPRIN-1 protein and is preferably breast cancer, kidney cancer, pancreatic cancer, large intestinal cancer, lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary 55 bladder cancer, esophageal cancer, leukemia, lymphoma, fibrosarcoma, mastocytoma, or melanoma.

Specific examples of these cancers include, but not limited to, breast adenocarcinoma, complex-type breast adenocarcinoma, malignant mixed tumor of mammary gland, intraductal papillary adenocarcinoma, lung adenocarcinoma, squamous cell cancer, small-cell cancer, large-cell cancer, glioma which is tumor of neuroepithelial tissue, ependymoma, neuronal tumor, embryonal neuroectodermal tumor, neurilemmoma, neurofibroma, meningioma, chronic lymphocytic leukemia, lymphoma, gastrointestinal lymphoma, alimentary lymphoma, small to medium cell-type lymphoma, cecal can-

cer, ascending colon cancer, descending colon cancer, transverse colon cancer, sigmoid colon cancer, rectal cancer, epithelial ovarian cancer, germ cell tumor, stromal cell tumor, pancreatic ductal carcinoma, invasive pancreatic ductal carcinoma, pancreatic adenocarcinoma, acinar cell carcinoma, adenosquamous carcinoma, giant cell tumor, intraductal papillary-mucinous neoplasm, mucinous cystic neoplasm, pancreatoblastoma, serous cystadenocarcinoma, solid-pseudopapillary tumor, gastrinoma, glucagonoma, insulinoma, multiple endocrine neoplasia type-1 (Wermer's syndrome), nonfunctional islet cell tumor, somatostatinoma, and VIPoma.

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The recipient subjects (patients) are preferably mammals, for example, mammals including primates, pet animals, livestock, and sport animals and are particularly preferably humans, dogs, and cats.

In the case of using the antibody of the present invention as a pharmaceutical composition, the pharmaceutical composition can be formulated by a method generally known to those skilled in the art. For example, the pharmaceutical composition can be used in the form of a parenteral injection of an aseptic solution or suspension with water or any other pharmaceutically acceptable liquid. For example, the pharmaceutical composition may be formulated with the antibody mixed in a unit dosage form required for generally accepted pharmaceutical practice, in appropriate combination with pharmacologically acceptable carriers or media, specifically, sterilized water, physiological saline, plant oil, an emulsifier, a suspending agent, a surfactant, a stabilizer, a flavoring agent, an excipient, a vehicle, a preservative, a binder, etc. The amount of the active ingredient in such a preparation is determined such that an appropriate dose within the prescribed range can be achieved.

an animal carrying a xenograft tissue inoculated with a cell line expressing CAPRIN-1 either spontaneously or after 35 according to conventional pharmaceutical practice using a transfection.

An aseptic composition for injection can be formulated according to conventional pharmaceutical practice using a vehicle such as injectable distilled water.

Examples of aqueous solutions for injection include physiological saline, isotonic solutions containing glucose and other adjuvants, for example, D-sorbitol, D-mannose, D-mannitol, and sodium chloride. These solutions may be used in combination with an appropriate solubilizer, for example, an alcohol (specifically, ethanol) or a polyalcohol (e.g., propylene glycol and polyethylene glycol), or a nonionic surfactant, for example, polysorbate 80TM or HCO-60.

Examples of oily solutions include sesame oil and soybean oil. These solutions may be used in combination with benzyl benzoate or benzyl alcohol as a solubilizer. The solutions may be further mixed with a buffer (e.g., a phosphate buffer solution and a sodium acetate buffer solution), a soothing agent (e.g., procaine hydrochloride), a stabilizer (e.g., benzyl alcohol and phenol), and an antioxidant. The injection solutions thus prepared are usually charged into appropriate ampules.

The pharmaceutical composition of the present invention is administered orally or parenterally, preferably parenterally. Specific examples of its dosage forms include injections, intranasal administration agents, transpulmonary administration agents, and percutaneous administration agents. Examples of the injections include intravenous injection, intramuscular injection, intraperitoneal injection, and subcutaneous injection, through which the pharmaceutical composition can be administered systemically or locally.

Also, the administration method can be appropriately selected depending on the age, weight, sex, symptoms, etc. of a patient. The dose of a pharmaceutical composition containing the antibody or a polynucleotide encoding the antibody can be selected within a range of, for example, 0.0001 to 1000 mg/kg of body weight per dose. Alternatively, the dose can be

selected within a range of, for example, 0.001 to 100000 mg/body of a patient, though the dose is not necessarily limited to these numeric values. Although the dose and the administration method vary depending on the weight, age, sex, symptoms, etc. of a patient, those skilled in the art can 5 appropriately select the dose and the method.

The pharmaceutical composition comprising the antibody of the present invention or the fragment thereof can be administered to a subject to treat and/or prevent cancer, preferably breast cancer, kidney cancer, pancreatic cancer, large intestinal cancer, lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary bladder cancer, esophageal cancer, leukemia, lymphoma, fibrosarcoma, mastocytoma, or melanoma.

The present invention further encompasses a method for 15 treating and/or preventing cancer, comprising administering the pharmaceutical composition of the present invention in combination with the antitumor agent as exemplified above or a pharmaceutical composition comprising the antitumor agent to a subject. The antibody of the present invention or the 20 fragment thereof may be administered simultaneously with or separately from the antitumor agent to the subject. In the case of separately administering these pharmaceutical compositions, either one may be administered first or later. Their dosing intervals, doses, administration routes, and the num- 25 ber of doses can be appropriately selected by a specialist. Other dosage forms of drugs to be administered simultaneously also include, for example, pharmaceutical compositions formulated by mixing the antibody of the present invention or the fragment thereof and the antitumor agent with a 30 pharmacologically acceptable carrier (or medium). The above descriptions about prescription, formulation, administration routes, doses, cancer, etc. as to the pharmaceutical compositions and dosage forms containing the antibody of the present invention are also applicable to any of the above- 35 described pharmaceutical compositions and dosage forms containing the antitumor agent.

Thus, the present invention also provides a combination drug for treatment and/or prevention of cancer, comprising the pharmaceutical composition of the present invention and 40 a pharmaceutical composition comprising the antitumor agent as exemplified above, and a method for treating and/or preventing cancer, comprising administering the combination drug. The present invention also provides a pharmaceutical composition for treatment and/or prevention of cancer, 45 comprising the antibody of the present invention or the fragment thereof and the antitumor agent together with a pharmacologically acceptable carrier.

<Polypeptide and DNA>

The present invention further provides a DNA encoding the antibody of the present invention or the fragment (antibodybinding fragment) thereof. Such a DNA may be a DNA encoding the heavy and/or light chains of the antibody or may be a DNA encoding the heavy and/or light chain variable regions of the antibody. Such a DNA may also be a DNA 55 encoding each or a combination of the complementarity determining regions of the antibody. Such a DNA includes, for example, a heavy chain variable region-encoding DNA comprising nucleotide sequences encoding the amino acid sequences of SEQ ID NOs: 8, 9, and 10 and a light chain 60 variable region-encoding DNA comprising nucleotide sequences encoding the amino acid sequences of SEQ ID NOs: 12, 13, and 14, in the case of the antibody (a).

The complementarity determining regions (CDRs) encoded by the DNA having these sequences serve as regions 65 that determine the specificity of the antibody. Sequences encoding the other regions (i.e., constant regions and frame-

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work regions) of the antibody may therefore be sequences derived from other antibodies. In this context, "other antibodies" also include antibodies derived from non-human organisms and are preferably those derived from humans from the viewpoint of reducing adverse reactions. Specifically, in the DNA described above, regions encoding each framework region and each constant region in the heavy and light chains preferably comprise nucleotide sequences encoding corresponding human antibody-derived amino acid sequences.

Further examples of the DNA encoding the antibody of the present invention include a heavy chain variable region-encoding DNA comprising a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 11, and a light chain variable region-encoding DNA comprising a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 15, in the case of the antibody (a). In this context, the nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 11 is, for example, the nucleotide sequence of SEQ ID NO: 16. The nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 15 is, for example, the nucleotide sequence of SEQ ID NO: 17. When such a DNA comprises a region encoding each constant region in the heavy and light chains, this region preferably comprises a nucleotide sequence encoding a corresponding human antibody-derived amino acid sequence (amino acid sequence of each constant region in the heavy and light chains).

These antibody DNAs can be obtained, for example, by the methods described above or the following method: first, total RNAs are prepared from hybridomas producing the antibody of the present invention using a commercially available RNA extraction kit, and cDNAs are synthesized using reverse transcriptase and random primers or the like. Subsequently, the antibody-encoding cDNAs are amplified by PCR using oligonucleotide primers for conserved sequences of each variable region in known mouse antibody heavy and light chain genes. Sequences encoding the constant regions can be obtained by the PCR amplification of known sequences. The nucleotide sequence of the DNA can be incorporated into a plasmid or a phage for sequencing, for example, and determined according to a routine method.

The present invention further provides the following polypeptides and DNAs related to the antibody (a) or (b):

- (i) a polypeptide comprising the amino acid sequence of SEQ ID NO: 11 or 21, and a DNA encoding the polypeptide (e.g., a DNA comprising the nucleotide sequence of SEQ ID NO: 16 or 26);
- (ii) a polypeptide comprising the amino acid sequence of SEQ ID NO: 15 or 25, and a DNA encoding the polypeptide (e.g., a DNA comprising the nucleotide sequence of SEQ ID NO: 17 or 27);
- (iii) a heavy chain CDR polypeptide selected from the group consisting of the amino acid sequences represented by SEQ ID NOs: 8, 9, and 10 and SEQ ID NOs: 18, 19, and 20, and a DNA encoding the polypeptide; and
- (iv) a light chain CDR polypeptide selected from the group consisting of the amino acid sequences represented by SEQ ID NOs: 12, 13, and 14 and SEQ ID NOs: 22, 23, and 24, and a DNA encoding the polypeptide.

These polypeptides and DNAs can be prepared using gene recombination techniques as described above.

<Summary of the Present Invention>

The aspects of the present invention described above are summarized below.

(1) An antibody or a fragment thereof which has immunological reactivity with a partial CAPRIN-1 polypeptide consisting of the amino acid sequence set forth in SEQ ID NO: 5

or an amino acid sequence having 80% or higher sequence identity to the amino acid sequence.

- (2) The antibody or fragment thereof according to (1), wherein the antibody or fragment thereof has cytotoxic activity against a cancer cell expressing a CAPRIN-1 protein.
- (3) The antibody or fragment thereof according to (1) or (2), wherein the antibody is a monoclonal antibody or a polyclonal antibody.
- (4) The antibody or fragment thereof according to any of (1) to (3), wherein the antibody is a human antibody, a humanized antibody, a chimeric antibody, a single-chain antibody, or a multispecific antibody.
- (5) The antibody or fragment thereof according to any of (1) to (4), wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity ¹⁵ determining regions of SEQ ID NOs: 8, 9, and 10 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the ²⁰ CAPRIN-1 protein.
- (6) The antibody or fragment thereof according to any of (1) to (4), wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 18, 19, and 20 (CDR1, ²⁵ CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 22, 23, and 24 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 protein.
- (7) The antibody or fragment thereof according to any of (1) to (6), wherein the antibody or fragment thereof is conjugated with an antitumor agent.
- (8) A pharmaceutical composition for treatment and/or prevention of cancer, comprising an antibody or fragment ³⁵ thereof according to any of (1) to (7) as an active ingredient.
- (9) The pharmaceutical composition according to (8), wherein the cancer is breast cancer, kidney cancer, pancreatic cancer, large intestinal cancer, lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary bladder cancer, esophageal cancer, leukemia, lymphoma, fibrosarcoma, mastocytoma, or melanoma.
- (10) A combination drug for treatment and/or prevention of cancer, comprising a pharmaceutical composition according to (8) or (9) and a pharmaceutical composition comprising an antitumor agent.
- (11) A DNA encoding an antibody or fragment thereof according to any of (1) to (6).
- (12) A method for treating and/or preventing cancer, comprising administering an antibody or fragment thereof ⁵⁰ according to any of (1) to (7), a pharmaceutical composition according to (8) or (9), or a combination drug according to (10) to a subject.

EXAMPLES

Hereinafter, the present invention will be described more specifically with reference to Examples. However, the scope of the present invention is not intended to be limited by these specific examples.

Example 1

Analysis of CAPRIN-1 Expression in Each Tissue

CAPRIN-1 gene expression in canine and human normal tissues and various cell lines was examined by RT-PCR

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according to Example 1(4) of WO2010/016526. As a result, its strong expression was seen in the testis among the healthy canine tissues, whereas the expression was seen in canine breast cancer and adenocarcinoma tissues. As a result of also confirming the expression in human tissues, the expression was confirmed only in the testis among normal tissues, as with the canine CAPRIN-1 gene. By contrast, the expression was detected in many types of cancer cell lines, including 8 human breast cancer cell lines (ZR75-1, MCF7; T47D, SK-10 BR-3, MDA-MB-157, BT-20, MDA-MB-231V, and MRK-nu-1) and 4 pancreatic cancer cell lines (Capan-2, MIAPaCa-2, Panc-1, and BxPc-3), among cancer cells. These results demonstrated that CAPRIN-1 is expressed in the breast cancer cell lines and the pancreatic cancer cell lines, though its
15 expression is not seen in normal tissues other than the testis.

Example 2

Preparation of Mouse Monoclonal Antibody Against CAPRIN-1

(1) Preparation of Mouse Monoclonal Antibody

100 μg of a human CAPRIN-1 protein having the amino acid sequence of SEQ ID NO: 2 as prepared in Example 3 of WO2010/016526 was mixed with an equal amount of MPL+ TDM adjuvant (manufactured by Sigma-Aldrich Corp.). This mixture was used as an antigen solution per mouse. The antigen solution was intraperitoneally administered to each 6-week-old Balb/c mouse (manufactured by Japan SLC, Inc.). Then, 7 boosters were performed every 1 week to complete immunization. Three days after the final shot, the spleen of each mouse was excised and ground between two sterilized glass slides. Procedures of washing with PBS(-) (manufactured by Nissui Pharmaceutical Co., Ltd.) and removing the supernatant by centrifugation at 0.1500 rpm for 10 minutes were repeated three times to obtain spleen cells. The obtained spleen cells were mixed with mouse myeloma cells SP2/0 (purchased from ATCC) at a ratio of 10:1. 200 µl of an RPMI1640 medium containing 10% FBS was heated to 37° C. and mixed with 800 µl of PEG1500 (manufactured by Boehringer Ingelheim GmbH), and the PEG solution thus prepared was added to the cell mixture, which was then left standing for 5 minutes for cell fusion. After removal of the supernatant by centrifugation at 1700 rpm for 5 minutes, the cells were suspended in 150 ml of an RPMI1640 medium containing 15% FBS supplemented with 2% equivalent of a HAT solution (manufactured by Life Technologies, Inc./ Gibco) (HAT selective medium). This suspension was added to fifteen 96-well plates (manufactured by Thermo Fisher Scientific Inc./Nunc) at 100 µl/well. The spleen cells and the myeloma cells were fused by culture at 37° C. for 7 days under conditions of 5% CO₂ to obtain hybridomas.

The prepared hybridomas were screened for the binding affinity of antibodies produced by the hybridomas against 55 CAPRIN-1 proteins as an index. A 1 μg/ml solution of the CAPRIN-1 proteins prepared by the approach described in Example 3 of WO2010/016526 was added to a 96-well plate at 100 μl/well and left standing at 4° C. for 18 hours. Each well was washed three times with PBS-T. Then, a 0.5% bovine serum albumin (BSA) solution (manufactured by Sigma-Aldrich Corp.) was added thereto at 400 μl/well and left standing at room temperature for 3 hours. The solution in each well was discarded, and each well was washed three times with 400 μl of PBS-T. Then, the culture supernatant of each hybridoma obtained above was added thereto at 100 μl/well and left standing at room temperature for 2 hours. Each well was washed three times with PBS-T. Then, HRP-

labeled anti-mouse IgG (H+L) antibodies (manufactured by Invitrogen Corp.) diluted 5000-fold with PBS were added thereto at 100 μ l/well and left standing at room temperature for 1 hour. Each well was washed three times with PBS-T. Then, a TMB substrate solution (manufactured by Thermo 5 Fisher Scientific Inc.) was added thereto at 100 μ l/well and left standing for 15 to 30 minutes to cause color reaction. After the color development, the reaction was terminated by the addition of 1 N sulfuric acid at 100 μ l/well. The absorbance was measured at 450 nm and 595 nm using an absorption spectrometer. As a result, several hybridomas producing antibodies having high absorbance were selected.

The selected hybridomas were added to a 96-well plate at a density of 0.5 cells/well and cultured in the plate. One week later, hybridomas forming single colonies in the wells were 15 observed. The cells in these wells were further cultured, and the cloned hybridomas were screened for the binding affinity of antibodies produced by the hybridomas against CAPRIN-1 proteins as an index. A 1 µg/ml solution of the CAPRIN-1 proteins prepared in Example 3 of WO2010/016526 was 20 added to a 96-well plate at 100 µl/well and left standing at 4° C. for 18 hours. Each well was washed three times with PBS-T. Then, a 0.5% BSA solution was added thereto at 400 μl/well and left standing at room temperature for 3 hours. The solution in each well was discarded, and each well was 25 washed three times with 400 µl of PBS-T. Then, the culture supernatant of each hybridoma obtained above was added thereto at 100 µl/well and left standing at room temperature for 2 hours. Each well was washed three times with PBS-T. Then, HRP-labeled anti-mouse IgG (H+L) antibodies (manu- 30 factured by Invitrogen Corp.) diluted 5000-fold with PBS were added thereto at 100 µl/well and left standing at room temperature for 1 hour. Each well was washed three times with PBS-T. Then, a TMB substrate solution (manufactured by Thermo Fisher Scientific Inc.) was added thereto at 100 35 μl/well and left standing for 15 to 30 minutes to cause color reaction. After the color development, the reaction was terminated by the addition of 1 N sulfuric acid at 100 µl/well. The absorbance was measured at 450 nm and 595 nm using an absorption spectrometer. As a result, 61 hybridoma lines pro- 40 ducing monoclonal antibodies reactive with CAPRIN-1 proteins were obtained.

Next, these monoclonal antibodies were screened for antibodies reactive with the surface of breast cancer cells expressing CAPRIN-1. Specifically, 10⁶ cells of a human breast 45 cancer cell line MDA-MB-231V were centrifuged in a 1.5-ml microcentrifuge tube. 100 ul of the culture supernatant of each hybridoma obtained above was added thereto and left standing for 1 hour on ice. After washing with PBS, FITClabeled goat anti-mouse IgG antibodies (manufactured by 50 Invitrogen Corp.) diluted 500-fold with PBS containing 0.1% FBS were added thereto and left standing for 1 hour on ice. After washing with PBS, the fluorescence intensity was measured using FACSCalibur (Becton, Dickinson and Company). On the other hand, the same operation as above was 55 performed using the serum of each untreated 6-week-old Balb/c mouse diluted 500-fold with a medium for hybridoma culture, instead of the antibodies, to prepare a control. As a result, two monoclonal antibodies (anti-CAPRIN-1 antibodies #1 and #2) having stronger fluorescence intensity than that 60 of the control, i.e., reactive with the surface of breast cancer cells, were selected.

(2) Identification of CAPRIN-1 Epitope Recognized by Anti-CAPRIN-1 Monoclonal Antibody #1

The cancer cell surface-reactive monoclonal antibodies 65 against CAPRIN-1 (anti-CAPRIN-1 antibodies #1 and #2) obtained in the paragraph (1) were used to identify a

CAPRIN-1 epitope region recognized thereby. 93 candidate peptides each consisting of 12 to 16 amino acids in the amino acid sequence of the human CAPRIN-1 protein were synthesized and each dissolved at a concentration of 1 mg/ml in DMSO.

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Each peptide was dissolved at a concentration of 30 $\mu g/m$ in a 0.1 M sodium carbonate buffer solution (pH 9.6). The solution was added at 100 $\mu l/well$ to a 96-well plate (manufactured by Thermo Fisher Scientific Inc./Nunc, product No.: 436006) and left standing overnight at 4° C. The solution in each well was discarded, and 10 mM ethanolamine/0.1 M sodium carbonate buffer solution (PH 9.6) was added thereto at 200 $\mu l/well$ and left standing at room temperature for 1 hour. Then, the solution in each well was discarded, and each well was washed twice with PBS containing 0.5% Tween 20 (PBST) to prepare a peptide-immobilized plate.

The cell culture supernatant containing the anti-CA-PRIN-1 antibody #1 was added at $50\,\mu$ l/well to each plate thus obtained. After shaking at room temperature for 1 hour, the solution in each well was discarded, and each well was washed three times with PBST. Next, a secondary antibody solution containing HRP-labeled anti-mouse IgG (manufactured by Invitrogen Corp.) antibodies diluted 3000- to 4000-fold with PBST was added thereto at $50\,\mu$ l/well. Then, the solution in each well was discarded, and each well was washed six times with PBST.

A TMB substrate solution (manufactured by Thermo Fisher Scientific Inc.) was added thereto at 100 μ l/well and left standing for 15 to 30 minutes to cause color reaction. After the color development, the reaction was terminated by the addition of 1 N sulfuric acid at 100 μ l/well. The absorbance was measured at 450 nm and 595 nm using an absorption spectrometer.

As a result, the anti-CAPRIN-1 antibodies #1 and #2 specifically reacted with a polypeptide of SEQ ID NO: 5. Of these 93 candidate peptides each consisting of 12to 16 amino acids, polypeptides of Asn Met Asn Ala Pro Val Pro Pro Val Asn Glu Pro (SEQ ID NO: 28) and Pro Val Pro Pro Val Asn Glu Pro Glu Thr Leu Lys Gln Gln Asn Gln (SEQ ID NO: 29), which partially overlapped with the polypeptide of SEQ ID NO: 5 in their amino acid sequences, did not respond to the anti-CAPRIN-1 antibodies #1 and #2obtained in Example 2. From these results, the polypeptide of SEQ ID NO: 5 was identified as a partial sequence of CAPRIN-1 specifically recognized by the anti-CAPRIN-1 antibodies #1 and #2obtained in Example 2(1).

(3) Cloning of Variable Region Genes of Anti-CAPRIN-1 Antibodies #1 and #2

The monoclonal antibodies obtained in Example 2(1) were analyzed for their variable region-encoding gene sequences and amino acid sequences thereof according to the method described in Example 5 of WO2010/016526. As a result, the monoclonal antibody #1 comprised a heavy chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 11 and a light chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 15. The antibody #2 comprised a heavy chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 21 and a light chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 25. A gene sequence encoding the heavy chain variable region of the obtained monoclonal antibody #1 is shown in SEQ ID NO: 16, and a gene sequence encoding the light chain variable region thereof is shown in SEQ ID NO: 17. A gene sequence encoding the heavy chain variable region of the antibody #2 is shown in SEQ ID NO: 26, and a gene sequence encoding the light chain variable region thereof is shown in SEQ ID NO 27.

It was also shown that: the monoclonal antibody #1 obtained in Example 2(1) comprises the heavy chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 11 and the light chain variable region consisting of the amino acid sequence set forth in SEO ID NO: 15, wherein CDR1, CDR2, and CDR3 in the heavy chain variable region consist of the amino acid sequences represented by SEQ ID NOs: 8, 9, and 10, respectively, and CDR1, CDR2, and CDR3 in the light chain variable region consist of the amino acid sequences represented by SEQ ID NOs: 12, 13, and 14, respectively; and the antibody #2 comprises the heavy chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 21 and the light chain variable region consisting of the amino acid sequence set forth in SEQ ID NO: 25, wherein CDR1, CDR2, and CDR3 in the heavy chain variable region consist of the amino acid sequences represented by SEQ ID NOs: 18, 19, and 20, respectively, and CDR1, CDR2, and CDR3 in the light chain variable region consist of the amino acid sequences represented by SEQ ID NOs: 22, 23, and 24, respectively.

Example 3

Preparation of Polyclonal Antibody Against Partial CAPRIN-1 Polypeptide Present on Cancer Cell Surface

In order to obtain polyclonal antibodies against partial CAPRIN-1 polypeptides present on cancer cell surface, a polypeptide (CAPRIN-1-derived peptide shown in SEQ ID 30 NO: 5) comprising the epitope region for the anti-CAPRIN-1 antibody #1 obtained in Example 2, a polypeptide having a region of amino acid residue numbers 50 to 98 in the human CAPRIN-1 amino acid sequence of SEQ ID NO: 2, and a polypeptide having a region of amino acid residue numbers 35 233 to 305 of SEQ ID NO: 2 were synthesized. 1 mg each of these peptides was mixed as an antigen with an equal volume of an incomplete Freund's adjuvant (IFA) solution. This mixture was subcutaneously administered to each rabbit four times every two weeks. Then, blood was collected to obtain 40 antiserum containing each polyclonal antibody. This antiserum was further purified using a protein G carrier (manufactured by GE Healthcare Bio-Sciences Ltd.) and replaced with PBS to obtain polyclonal antibodies against partial addition, the serum of a rabbit that received no antigen was purified using a protein G carrier in the same way as above and used as control antibodies.

Example 4

Analysis of CAPRIN-1 Protein Expression on Cancer Cell Membrane Surface

Next, 8 human breast cancer cell lines (ZR75-1, MCF7, 55 T47D, SK-BR-3, MDA-MB-157, BT-20, MDA-MB-231V, and MRK-nu-1) confirmed to have a large level of CAPRIN-1 gene expression were examined for their expression of CAPRIN-1 proteins on the cell surface. 5×10⁵ cells of each human breast cancer cell line thus confirmed to have gene 60 expression were centrifuged in a 1.5-ml microcentrifuge tube. 2 μg (5 μl) each of the polyclonal antibodies against CAPRIN-1-derived peptides represented by SEQ ID NO: 5 prepared as described above in Example 3 and 95 µl of PBS containing 0.1% fetal bovine serum were added thereto and mixed, and left standing for 1 hour on ice. After washing with PBS, the resulting solution was mixed by the addition of 1 μ l

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of Alexa 488-labeled goat anti-rabbit IgG antibodies (manufactured by Invitrogen Corp.) and 98 µl of PBS containing 0.1% fetal bovine serum (FBS) and left standing for 30 hours on ice. After washing with PBS, the fluorescence intensity was measured using FACSCalibur (Becton, Dickinson and Company). On the other hand, the same operation as above was performed using the control antibodies prepared as described above in Example 3 instead of the polyclonal antibodies against CAPRIN-1-derived peptides to prepare a control. As a result, the cancer cells supplemented with the anti-CAPRIN-1 antibodies all exhibited fluorescence intensity at least 35% stronger than that of the control. This demonstrated that CAPRIN-1 proteins are expressed on the cell membrane surface of the human cancer cell lines. The above rate of enhancement in fluorescence intensity was indicated by the rate of increase in mean fluorescence intensity (MFI) in each cell line and calculated according to the following expression:

Rate of increase in mean fluorescence intensity(Rate of enhancement in fluorescence intensity)(%)= ((MFI of cells reacted with the anti-CAPRIN-1 antibodies)-(Control MFI))/(Control MFI)×100

Also, the fluorescence intensity was measured in 2 kidney cancer cell lines (Caki-1 and Caki-2), a urinary bladder cancer cell line (T24), an ovary cancer cell line (SKOV3), 2 lung 25 cancer cell lines (QG56 and A549), a prostate cancer cell line (PC3), a uterine cervix cancer cell line (SW756), a fibrosarcoma cell line (HT1080), 2 brain tumor cell lines (T98G and U87MG), a gastric cancer cell line (MNK28), 3 large intestinal cancer cell lines (Lovo, DLD-1, and HCT-116), and 4 pancreatic cancer cell lines (Capan-2, MIAPaCa-2, Panc-1, and BxPC-3) using the same approach as above. As a result, all the cancer cells had fluorescence intensity at least 35% stronger than that of the control.

As with the results obtained above, CAPRIN-1 protein expression on cancer cell membrane surface was also confirmed using the anti-CAPRIN-1 antibody #1 obtained in Example 2.

Example 5

Preparation of Human-mouse Chimeric Monoclonal Antibody

The gene amplification fragment comprising the gene of CAPRIN-1 polypeptides present on cancer cell surface. In 45 the heavy chain variable region of the anti-CAPRIN-1 antibody #1 or #2 obtained in Example 2 was treated at both ends with restriction enzymes, then purified, and inserted according to a routine method into a pcDNA4/myc-His (manufactured by Invitrogen Corp.) vector already having gene inserts 50 of a mouse antibody-derived leader sequence and a human IgG₁ H chain constant region comprising the amino acid sequence of SEQ ID NO: 6. Also, the gene amplification fragment comprising the gene of the light chain variable region of the anti-CAPRIN-1 antibody #1 was treated at both ends with restriction enzymes, then purified, and inserted according to a routine method into a pcDNA3.1/myc-His (manufactured by Invitrogen Corp.) vector already having gene inserts of a mouse antibody-derived leader sequence and a human IgG₁ L chain constant region comprising the amino acid sequence of SEQ ID NO: 7.

Next, the recombinant vector having the gene insert of the heavy chain variable region of the anti-CAPRIN-1 antibody #1 or #2 and the recombinant vector having the gene insert of the light chain variable region were introduced into CHO-K1 cells (obtained from Riken Cell Bank). Specifically, 2×10⁵ CHO-K1 cells were cultured in 1 ml of a Ham's F12 medium (manufactured by Invitrogen Corp.) containing 10% FBS per

well of a 12-well culture plate, and washed with PBS(-). Then, 1 ml of a fresh Ham's F12 medium containing 10% FBS per well was added thereto. 250 ng each of the vectors lysed in 30 µl of OptiMEM (manufactured by Invitrogen Corp.) was mixed with 30 µl of Polyfect transfection reagent 5 (manufactured by Qiagen N.V.), and this mixture was added to each well. The CHO-K1 cells cotransfected with the recombinant vectors were cultured in a Ham's F12 medium containing 10% FBS supplemented with 200 µg/ml Zeocin (manufactured by Invitrogen Corp.) and 200 µg/m Geneticin 10 (manufactured by Roche Diagnostics K.K.) and then inoculated to a 96-well plate at a density of 0.5 cells/well to prepare cell lines stably producing any of human-mouse chimeric monoclonal antibodies #1 and #2 having the variable regions of the anti-CAPRIN-1 antibodies #1 and #2, respectively, 15 obtained in Example 2.

Each prepared cell line was cultured for 5 days in a 150-cm² flask at a density of 5×10⁵ cells/ml using 30 ml of a serum-free OptiCHO medium (manufactured by Invitrogen Corp.) to obtain culture supernatants containing the humanmouse chimeric monoclonal antibody #1 or #2.

Also, cell lines stably producing human-mouse chimeric comparative antibodies 1 to 26 were prepared as comparative samples in the same way as above respectively using the following comparative antibodies: anti-CAPRIN-1 mouse- 25 derived monoclonal antibodies described in WO2010/ 016526 [a comparative antibody 1 having the heavy chain variable region of SEQ ID NO: 26 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 27; a comparative antibody 2 30 having the heavy chain variable region of SEQ ID NO: 28 and the light chain variable region of SEQ ID NO: 29; a comparative antibody 3 having the heavy chain variable region of SEQ ID NO: 30 and the light chain variable region of SEQ ID NO: 31; a comparative antibody 4 having the heavy chain variable 35 region of SEQ ID NO: 32 and the light chain variable region of SEQ ID NO: 33; a comparative antibody 5 having the heavy chain variable region of SEQ. ID NO: 34 and the light chain variable region of SEQ ID NO: 35; a comparative antibody 6 having the heavy chain variable region of SEQ ID 40 NO: 36 and the light chain variable region of SEQ ID NO: 37; a comparative antibody 7 having the heavy chain variable region of SEQ ID NO: 38 and the light chain variable region of SEQ ID NO: 39; a comparative antibody 8 having the heavy chain variable region of SEQ ID NO: 40 and the light 45 chain variable region of SEQ ID NO: 41; a comparative antibody 9 having the heavy chain variable region of SEO ID NO: 42 and the light chain variable region of SEQ ID NO: 43; a comparative antibody 10 having the heavy chain variable region of SEQ ID NO: 44 and the light chain variable region 50 of SEQ ID NO: 45; and a comparative antibody 11 having the heavy chain variable region of SEQ ID NO: 46 and the light chain variable region of SEQ ID NO: 47], anti-CAPRIN-1 monoclonal antibodies described in WO2011/096517 [a comparative antibody 12 having the heavy chain variable 55 region of SEQ ID NO: 43 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 47; and a comparative antibody 13 having the heavy chain variable region of SEQ ID NO: 43 and the light chain variable region of SEQ ID NO:], anti-CA- 60 PRIN-1 monoclonal antibodies described in WO2011/ 096528 [a comparative antibody 14 having the heavy chain variable region of SEQ ID NO: 43 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 47; a comparative antibody 15 having the heavy chain variable region of SEQ ID NO: 51 and the light chain variable region of SEQ ID NO: 55; a compara30

tive antibody 16 having the heavy chain variable region of SEQ ID NO: 59 and the light chain variable region of SEQ ID NO: 63; a comparative antibody 17 having the heavy chain variable region of SEQ ID NO: 76 and the light chain variable region of SEQ ID NO: 80; a comparative antibody 18 having the heavy chain variable region of SEQ ID NO: 84 and the light chain variable region of SEQ ID NO: 88; and a comparative antibody 19 having the heavy chain variable region of SEQ ID NO: 92 and the light chain variable region of SEQ ID NO: 96], an anti-CAPRIN-1 monoclonal antibody described in WO2011/096519 [a comparative antibody 20 having the heavy chain variable region of SEQ ID NO: 42 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 46], anti-CAPRIN-1 monoclonal antibodies described in WO2011/096533 [a comparative antibody 21 having the heavy chain variable region of SEQ ID NO: 43 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 51; a comparative antibody 22 having the heavy chain variable region of SEQ ID NO: 47 and the light chain variable region of SEQ ID NO: 51; and a comparative antibody 23 having the heavy chain variable region of SEQ ID NO: 63 and the light chain variable region of SEQ ID NO: 67], and anti-CAPRIN-1 monoclonal antibodies described in WO2011/096534 [a comparative antibody 24 having the heavy chain variable region of SEQ ID NO: 43 (described therein; the same holds true for the description below) and the light chain variable region of SEQ ID NO: 47; a comparative antibody 25 having the heavy chain variable region of SEQ ID NO: 43 and the light chain variable region of SEQ ID NO: 51; and a comparative antibody 26 having the heavy chain variable region of SEQ ID NO: 63 and the light chain variable region of SEQ ID NO: 67]. Each prepared cell line was cultured for 5 days in a 150-cm² flask at a density of 5×10⁵ cells/ml using 30 ml of a serum-free OptiCHO medium (manufactured by Invitrogen Corp.) to obtain culture supernatants containing any of the humanmouse chimeric comparative monoclonal antibodies 1 to 26.

Example 6

Expression of CAPRIN-1 on Surface of Various Cancer Cells Using Anti-CAPRIN-1 Monoclonal Antibody

Next, the 8 human breast cancer cell lines (ZR75-1, MCF7, T47D, SK-BR-3, MDA-MB-157, BT-20, MDA-MB-231V, and MRK-nu-1), the 2 kidney cancer cell lines (Caki-1 and Caki-2), the urinary bladder cancer cell line (T24), the ovary cancer cell line (SKOV3), the 2 lung cancer cell lines (QG56 and A549), the prostate cancer cell line (PC3), the uterine cervix cancer cell line (SW756), the fibrosarcoma cell line (HT1080), the 2 brain tumor cell lines (T98G and U87MG), the gastric cancer cell line (MNK28), the 3 large intestinal cancer cell lines (Lovo, DLD-1, and HCT-116), and the 4 pancreatic cancer cell lines (Capan-2, MIAPaCa-2, Panc-1, and BxPC-3) confirmed to have CAPRIN-1 gene expression were examined for their expression of CAPRIN-1 proteins on the cell surface using the culture supernatants respectively containing the anti-CAPRIN-1 antibodies #1 and #2 obtained in Example 2. 10⁶ cells of each cell line were centrifuged in each 1.5-ml microcentrifuge tube. Each culture supernatant (100 µl) containing the antibody was added to the tube and left standing for 1 hour on ice. After washing with PBS, FITClabeled goat anti-mouse IgG (H+L) antibodies (manufactured by Jackson ImmunoResearch Laboratories, Inc.) diluted with PBS containing 0.1% FBS were added thereto and left stand-

ing at 4° C. for 30 minutes. After washing with PBS, the fluorescence intensity was measured using FACSCalibur (Becton, Dickinson and Company). The negative control used was cells reacted only with secondary antibodies. As a result, the anti-CAPRIN-1 antibodies #1 and #2 each exhibited reactivity with fluorescence intensity at least 30% stronger than that of the negative control. This demonstrated that CAPRIN-1 proteins are expressed on the cell membrane surface of the human cancer cell lines. The above rate of enhancement in fluorescence intensity was indicated by the 10 rate of increase in mean fluorescence intensity (MFI) in each cell line and calculated according to the following expression:

Rate of increase in mean fluorescence intensity(Rate of enhancement in fluorescence intensity)(%)= ((MFI of cells reacted with the anti-CAPRIN-1 antibodies)–(Control MFI))/(Control MFI)×100

Example 7

Antitumor Activity Against Cancer Cell of Antibody Against CAPRIN-1-Derived Peptide (SEQ ID NO:

In order to evaluate each antibody against the CAPRIN-1-derived peptide represented by SEQ ID NO: 5 for the strength 25 of its cytotoxicity against cancer cells expressing CAPRIN-1, ADCC activity was determined. The polyclonal antibodies against the peptide represented by SEQ ID NO: 5 prepared in Example 3 were used in this evaluation. Similar evaluation was conducted using polyclonal antibodies against other 30 human CAPRIN-1-derived peptides (polyclonal antibodies against amino acid residue numbers 50 to 98 in the amino acid sequence of human CAPRIN-1 and polyclonal antibodies against amino acid residue numbers 233 to 305, which were prepared in Example 3) as antibodies to be compared and the 35 rabbit serum-derived control antibodies prepared in Example 3 as a negative control.

10⁶ cells each of the human breast cancer cell line MDA-MB-231V, the human large intestinal cancer cell line DLD-1, the human pancreatic cancer cell line Capan-2, and the human 40 lung cancer cell line QG56 confirmed to have CAPRIN-1 expression were collected into a 50-ml centrifuge tube, to which 100 μCi of chromium 51 was then added, followed by incubation at 37° C. for 2 hours. Then, the cells were washed three times with an RPMI1640 medium containing 10% fetal 45 calf serum and added at a density of 2×10^3 cells/well to each 96-well V-bottom plate. The polyclonal antibodies against the human CAPRIN-1-derived peptide represented by SEQ ID NO: 5 and two types of polyclonal antibodies against other human CAPRIN-1-derived peptides (polyclonal antibodies 50 against amino acid residue numbers 50 to 98 in SEQ ID NO: 2 of human CAPRIN-1 and polyclonal antibodies against amino acid residue numbers 233 to 305) as described above were separately added thereto at a concentration of 1 μg/well. Lymphocytes separated from human or rabbit peripheral 55 blood according to a routine method were further added thereto at a density of 4×10^5 cells/well and cultured at 37° C. for 4 hours under conditions of 5% CO₂. After the culture, the amount of chromium (Cr) 51 released from damaged cancer cells was measured in the culture supernatant to calculate the 60 ADCC activity against the cancer cells of the polyclonal antibodies against each human CAPRIN-1-derived peptide. As a result, all the polyclonal antibodies obtained by immunization with the partial peptides of human CAPRIN-1 having an amino acid sequence of amino acid residue numbers 50 65 to 98 or amino acid residue numbers 233 to 305 of SEQ ID NO: 2 of human CAPRIN-1 had activity less than 8% against

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the human breast cancer cell line MDA-MB-231V, the human large intestinal cancer cell line DLD-1, the human pancreatic cancer cell line Capan-2, and the human lung cancer cell line QG56. By contrast, the groups supplemented with the polyclonal antibodies against the human CAPRIN-1-derived peptide represented by SEQ ID NO: 5 were confirmed to have 27% or higher cytotoxic activity against all the cancer cell lines. The negative control antibodies had activity less than 5% against all the cancer cells. These results demonstrated that the antibody against CAPRIN-1 shown in SEQ ID NO: 5 exerts strong cytotoxic activity against cancer cells expressing CAPRIN-1.

These results about cytotoxic activity were obtained by: mixing the antibody against CAPRIN-1 used in the present invention, lymphocytes, and 2×10³ cells of each cancer cell line with incorporated chromium 51, as described above: culturing the cells for 4 hours; after the culture, measuring the amount of chromium 51 released into the medium; and calculating the cytotoxic activity against each cancer cell line according to the following expression*:

Cytotoxic activity(%)=Amount of chromium51 released from the target cells supplemented with the antibody against CAPRIN-1 and lymphocytes/Amount of chromium 51 released from target cells supplemented with 1 N hydrochloric acidx100

*Expression

The human-mouse chimeric monoclonal antibodies #1 and #2 obtained in Example 5 were evaluated for their cytotoxic activity against human cancer cells. The culture supernatant of each cell line producing any of the antibodies was purified using Hitrap Protein A Sepharose FF (manufactured by GE Healthcare Bio-Sciences Ltd.). After replacement with PBS (-), the solution was filtered through a 0.22-μm filter (manufactured by Millipore Corp.). The resulting antibody was used for activity assay. 10° cells each of the human breast cancer cell line MDA-MB-231V, the human large intestinal cancer cell line DLD-1, the human pancreatic cancer cell line Capan-2, and the human lung cancer cell line QG56 were collected into a 50-ml centrifuge tube, to which 100 µCi of chromium 51 was then added, followed by incubation at 37° C. for 2 hours. Then, the cells were washed three times with an RPMI1640 medium containing 10% FBS and added at a density of 2×10³ cells/well to each 96-well V-bottom plate to prepare target cells. The purified antibodies (human-mouse chimeric monoclonal antibodies #1 and #2) and the humanmouse chimeric comparative monoclonal antibodies 1 to 26 obtained in Example 5 were each added thereto at a concentration of 0.75 µg/well. A cell population containing human NK cells was separated using a routine method from human peripheral blood lymphocytes prepared according to a routine method. The cell population containing human NK cells that was used in this evaluation was prepared as follows: human peripheral blood mononuclear cells separated using a specific gravity separation solution Histopaque for peripheral blood mononuclear cell separation (Sigma-Aldrich Corp.) were reacted with FITC fluorescent dye-labeled antibodies (antihuman CD3 antibody, anti-human CD20 antibody, anti-human CD19 antibody, anti-human CD11c antibody, or anti-HLA-DR antibody (Becton, and Dickinson and Company)), and a cell population containing NK cells unstained with the antibodies was separated using a cell sorter (FACS Vantage SE (Becton, and Dickinson and Company)) or human NK cell separation kit (manufactured by Miltenyi Biotec K.K.). The separated cell population containing NK cells was added to the plate at a density of 2×10^5 cells/well and cultured at 37° C. for 4 hours under conditions of 5% CO₂. After the culture, the amount of chromium 51 released from damaged tumor cells was measured in the culture supernatant to calculate the cytotoxic activity of each anti-CAPRIN-1 antibody against the

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cancer cells. The negative control used was cells supplemented with isotype control antibodies. As a result, the isotype control antibodies used had cytotoxic activity of less than 5% against all of the cancer cell lines, and the humanmouse chimeric comparative monoclonal antibodies 1 to 26 used had cytotoxic activity of less than 5% against MDA-MB-231V, less than 8% against DLD-1, less than 6% against Capan-2, and less than 6% against QG56. By contrast, the human-mouse chimeric anti-CAPRIN-1 antibody #1 had cytotoxic activity of 15% or higher against MDA-MB-231V, 24% or higher against DLD-1, 26% or higher against Capan-2, and 19% or higher against QG56. Likewise, the isotype control antibodies used and the comparative antibodies 1 to 26 used had cytotoxic activity less than 4% against all of other cancer cells, breast cancer cell lines T47D, Hs578T, BT-20, SK-BR-3, MCF7, and MRK-nu-1, a glioma cell line T98G, a lung cancer cell line A549, a kidney cancer cell line Caki-1, a uterine cervix cancer cell line SW756, a urinary bladder cancer cell line T24, a gastric cancer cell line MKN28, a large intestinal cancer cell line SW480, a leukemia cell line AML5, and a lymphoma cell line Ramos. By contrast, the humanmouse chimeric monoclonal antibodies #1 and #2 were con- 20 firmed to have 10% or higher cytotoxic activity against these cell lines. These results showed that the antibodies against the CAPRIN-1-derived peptide shown in SEQ ID NO: 5 damage CAPRIN-1-expressing cancer cells through their ADCC activity, and demonstrated that the human-mouse chimeric monoclonal antibodies #1 and #2 exhibit stronger cytotoxic 25 activity against human cancer cells than that of the comparative antibodies 1 to 26.

These results about cytotoxic activity were obtained by: mixing the antibody against CAPRIN-1 used in the present invention, lymphocytes (cell population containing NK $_{30}$ cells), and 2×10^3 cells of each cancer cell line with incorporated chromium $_{51}$, as described above: culturing the cells for 4 hours; after the culture, measuring the amount of chromium $_{51}$ released into the medium; and calculating the cytotoxic activity against each cancer cell line according to the following expression*:

Cytotoxic activity(%)=Amount of chromium 51 released from the target cells supplemented with the antibody against CAPRIN-1 and lymphocytes (cell population containing NK cells)/Amount of chromium 51 released from target cells supplemented with 1 N hydrochloric acidx100.

*Expression

Example 8

The Number of CAPRIN-1 Molecules on Surface of Various Cancer Cells Recognized by Anti-CAPRIN-1 Antibodies #1 and #2

A human breast cancer cell line (MDA-MB-231V), a kidney cancer cell line (Caki-1), a urinary bladder cancer cell line

(T24), an ovary cancer cell line (SKOV3), lung cancer cell lines (QG56 and A549), a pancreatic cancer cell line (Capan-2), a prostate cancer cell line (PC3), a uterine cervix cancer cell line (SW756), a fibrosarcoma cell line (HT1080), a brain tumor cell line (T98G), a gastric cancer cell line (MKN28), large intestinal cancer cell lines (Lovo and DLD-1), a leukemia cell line (AML5), and a lymphoma cell line (Ramos) were examined using an assay kit "QIFIKIT" for the number of molecules (manufactured by Dako Japan Inc.) for the number of CAPRIN-1 molecules on their cell surface recognized by the anti-CAPRIN-1 antibodies #1 and #2. Similarly, the number of CAPRIN-1 molecules on the surface of these various cancer cells was also examined using the anti-CAPRIN-1 comparative monoclonal antibodies 1 to 26 prepared in Example 5.

According to the protocol attached to the kit, each antibody (anti-CAPRIN-1 antibodies #1 and comparative antibodies 1 to 26) was diluted into 5 µg/ml (in terms of final concentration) with PBS, and this dilution was added to each cell line and reacted for 30 minutes. After washing with PBS, fluorescently labeled anti-mouse IgG antibodies attached to the kit were added as secondary antibodies, together with calibration beads attached to the kit, to each cell line and left standing for 45 minutes on ice. Each cell line and the calibration beads were washed with PBS. Then, the fluorescence intensity was measured using FACSCalibur (Becton, Dickinson and Company) to obtain a mean fluorescence intensity value (mean). Also, a mean fluorescence intensity value (mean) was obtained by the same assay as above for the comparative antibodies. The negative control used was cells reacted with isotype control antibodies, and a mean was also obtained. Each mean fluorescence intensity value (mean) was used to calculate the number of molecules according to the protocol attached to the kit. As a result, the number of CAPRIN-1 molecules on the surface of various cancer cells recognized by the anti-CAPRIN-1 monoclonal antibodies #1 and #2 and the comparative antibodies 12 to 26 was 10⁵ or more per cell for all the examined human cancer cell lines. On the other hand, the number of molecules recognized by the comparative antibodies 1 to 11 was less than 10⁵ per cell.

INDUSTRIAL APPLICABILITY

The antibody of the present invention is useful for the treatment and/or prevention of cancer.

All publications, patents, and patent applications cited herein are incorporated herein by reference in their entirety.

SEQUENCE LISTING

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Gly Ala Ala Ala Pro Ala Ser Gln His Pro Ala Thr Gly Thr Gly Ala 35 40 45	
Val Gln Thr Glu Ala Met Lys Gln Ile Leu Gly Val Ile Asp Lys Lys 50 55 60	

Leu Arg Asn Leu Glu Lys Lys Lys Gly Lys Leu Asp Asp Tyr Gln Glu 65 70 75 80

Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp Ala Val\$85\$ 90 95

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Gln	Arg	Ser 115	Phe	Met	Ala	Leu	Ser 120	Gln	Asp	Ile	Gln	Lys 125	Thr	Ile	Lys
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Arg 145	Leu	Lys	Thr	Val	Leu 150	Glu	Leu	Gln	Tyr	Val 155	Leu	Asp	rys	Leu	Gly 160
Asp	Asp	Glu	Val	Arg 165	Thr	Asp	Leu	Lys	Gln 170	Gly	Leu	Asn	Gly	Val 175	Pro
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His	Ala 210	Ser	Ile	His	Leu	Trp 215	Aap	Leu	Leu	Glu	Gly 220	ГÀа	Glu	ГЛа	Pro
Val 225	Cys	Gly	Thr	Thr	Tyr 230	TÀa	Val	Leu	_	Glu 235	Ile	Val	Glu	Arg	Val 240
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Tyr	Gln 450	Pro	Ser	His	Ala	Thr 455	Glu	Gln	Arg	Pro	Gln 460	Lys	Glu	Pro	Ile
Asp 465	Gln	Ile	Gln	Ala	Thr 470	Ile	Ser	Leu	Asn	Thr 475	Asp	Gln	Thr	Thr	Ala 480
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Asn Phe Lys Arg Gly Ser Gly Gln Ser Gly Pro Arg Gly Ala Pro Arg	
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cag gaa cga atg aac aaa ggg gaa agg ctt aat caa gat cag ctg gat Gln Glu Arg Met Asn Lys Gly Glu Arg Leu Asn Gln Asp Gln Leu Asp	471

						47					CD	·,_	,	,,,		48	
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	80					85					90						
-	_		_		_	_	_				_	gag Glu		_		519	
_		_		_		_	_		_		_	att Ile	_			567	
												gaa Glu				615	
												gtt Val 155				663	
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												aac Asn				951	
												gca Ala				999	
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									cag Gln							1863
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Ser	Lys	Tyr	Gln 100	Glu	Val	Thr	Asn	Asn 105	Leu	Glu	Phe	Ala	Lys 110	Glu	Leu	
Gln	Arg	Ser 115	Phe	Met	Ala	Leu	Ser 120	Gln	Asp	Ile	Gln	Lys 125	Thr	Ile	Lys	
Lys	Thr 130	Ala	Arg	Arg	Glu	Gln 135	Leu	Met	Arg	Glu	Glu 140	Ala	Glu	Gln	Lys	

Asp Asp Glu Val Arg Thr Asp Leu Lys Gln Gly Leu Asn Gly Val Pro \$165\$

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His	Ala 210	Ser	Ile	His	Leu	Trp 215	Asp	Leu	Leu	Glu	Gly 220	Lys	Glu	Lys	Pro
Val 225	Сув	Gly	Thr	Thr	Tyr 230	Lys	Val	Leu	Lys	Glu 235	Ile	Val	Glu	Arg	Val 240
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Cys	Glu	Glu	Glu 260	Glu	Ala	Ala	Ser	Ala 265	Pro	Ala	Val	Glu	Asp 270	Gln	Val
Pro	Glu	Ala 275	Glu	Pro	Glu	Pro	Ala 280	Glu	Glu	Tyr	Thr	Glu 285	Gln	Ser	Glu
Val	Glu 290	Ser	Thr	Glu	Tyr	Val 295	Asn	Arg	Gln	Phe	Met 300	Ala	Glu	Thr	Gln
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Val	Glu	Val	Val	Asn 325	Ser	Leu	Gln	Gln	Gln 330	Pro	Gln	Ala	Ala	Ser 335	Pro
Ser	Val	Pro	Glu 340	Pro	His	Ser	Leu	Thr 345	Pro	Val	Ala	Gln	Ala 350	Asp	Pro
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Pro	Tyr 370	Asn	Phe	Ile	Gln	Asp 375	Ser	Met	Leu	Asp	Phe 380	Glu	Asn	Gln	Thr
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Met	Asp	Met	Pro	Gln 405	Leu	Val	Cha	Pro	Pro 410	Val	His	Ser	Glu	Ser 415	Arg
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Pro	Leu	Val 435	Ser	Ser	Thr	Ser	Glu 440	Gly	Tyr	Thr	Ala	Ser 445	Gln	Pro	Leu
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Phe	Gln	Ser 515	Met	Gln	Thr	Val	Phe 520	Asn	Met	Asn	Ala	Pro 525	Val	Pro	Pro
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Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
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Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser Thr
Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys
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Asn Asn Tyr Ala Thr Tyr Tyr Ala Asp Ser Val Lys Asp Arg Phe Thr 50 \\ 60
Ile Ser Arg Asp Asp Ser Gln Ser Met Leu Tyr Leu Gln Met Asn Asn 65 70 75 80
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Met His Trp Asn Gln Gln Lys Pro Gly Gln Pro Pro Arg Leu Leu Ile
Tyr Leu Val Ser Asn Leu Glu Ser Gly Val Pro Ala Arg Phe Ser Gly
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gttgctcgca taagaagtaa aagtaataat tatgcaacat attatgccga ttcagtgaaa
                                                                      180
gacaggttca ccatctccag agatgattca caaagcatgc tctatctgca aatgaacaac
                                                                      240
ttgaaaactg aggacacagc catgtattac tgtgtgagag attgggatgg tttcctttac
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Arg Pro Gly Gln Gly Leu Glu Trp Ile Gly Met Ile Asp Pro Ser Asp 35 40 45
Ser Glu Thr Arg Leu Asn Gln Lys Phe Lys Asp Lys Ala Thr Leu Thr
                      55
Val Asp Lys Ser Ser Ser Thr Ala Tyr Met Gln Leu Ser Ser Pro Thr 65 70 75 80
Ser Glu Asp Ser Ala Val Tyr Tyr Cys Ala Thr Tyr Pro Asp Trp Ala
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Lys Ala His Ser Pro Leu Arg
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Leu Asn Trp Phe Leu Gln Arg Pro Gly Gln Ser Pro Gln Leu Leu Ile
Tyr Leu Met Ser Thr Arg Ala Ser Gly Val Ser Asp Arg Phe Ser Gly
Ser Gly Ser Gly Thr Asp Phe Thr Leu Glu Ile Ser Arg Val Lys Ala
Glu Asp Val Gly Val Tyr Tyr Cys Gln Gln Leu Val Glu Asp Pro Leu
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attggcatga ttgatccttc cgatagtgaa actaggttaa atcagaagtt caaggacaag
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gccacattga ctgtagacaa atcctccagc acagcctaca tgcaactcag cagcccgaca
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cggtttagtg gcagtgggtc aggaacagat ttcaccctgg aaatcagtag agtgaaggct
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qaqqatqtqq qtqtqtatta ctqtcaacaa cttqtaqaqq atccqctcac qttcqqtqct
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The invention claimed is:

- 1. An antibody or a fragment thereof which has immunological reactivity with a partial CAPRIN-1 polypeptide consisting of the amino acid sequence set forth in SEQ ID NO: 5.
- 2. The antibody or fragment thereof according to claim 1, wherein the antibody or fragment thereof has cytotoxic activity against a cancer cell expressing a CAPRIN-1protein.
- 3. The antibody or fragment thereof according to claim 1, 20 wherein the antibody is a monoclonal antibody or a polyclonal antibody.
- 4. The antibody or fragment thereof according to claim 1, wherein the antibody is a human antibody, a humanized antibody, a chimeric antibody, a single-chain antibody, or a mul- 25 tispecific antibody.
- 5. The antibody or fragment thereof according to claim 1, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 8, 9, and 10 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 6. The antibody or fragment thereof according to claim 1, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 18, 19, and 20 (CDR1, CDR2, 40 and CD3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 22, 23, and 24 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 7. The antibody or fragment thereof according to claim 1, wherein the antibody or fragment thereof is conjugated with an antitumor agent.
- 8. A pharmaceutical composition for treatment of of a caprin-1-expressing cancer, comprising an antibody or fragment thereof according to claim 1 as an active ingredient.
- 9. The pharmaceutical composition according to claim 8, wherein the cancer is breast cancer, kidney cancer, pancreatic cancer, large intestinal cancer, lung cancer, brain tumor, gastric cancer, uterine cervix cancer, ovary cancer, prostate cancer, urinary bladder cancer, esophageal cancer, leukemia, lymphoma, fibrosarcoma, mastocytoma, or melanoma.
- 10. A combination drug for treatment of a caprin-1-expressing cancer, comprising a pharmaceutical composition 60 according to claim 8 and a pharmaceutical composition comprising an antitumor agent.
- 11. A DNA encoding an antibody or fragment thereof according to claim 1.
- 12. A method for treating a caprin-1-expressing cancer, 65 comprising administering an antibody or fragment thereof according to claim 1 to a subject.

13. The antibody or fragment thereof according to claim 2, wherein the antibody is a monoclonal antibody or a polyclonal antibody.

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- 14. The antibody or fragment thereof according to claim 2, wherein the antibody is a human antibody, a humanized antibody, a chimeric antibody, a single-chain antibody, or a multispecific antibody.
- 15. The antibody or fragment thereof according to claim 3, wherein the antibody is a human antibody, a humanized antibody, a chimeric antibody, a single-chain antibody, or a multispecific antibody.
- 16. The antibody or fragment thereof according to claim 2, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 8, 9, and 10 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 17. The antibody or fragment thereof according to claim 3, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 8, 9, and 10 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 18. The antibody or fragment thereof according to claim 4, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 8, 9, and 10 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 12, 13, and 14 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 19. The antibody or fragment thereof according to claim 2, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 18, 19, and 20 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID NOs: 22, 23, and 24 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 pro-
- 20. The antibody or fragment thereof according to claim 3, wherein the antibody or fragment thereof comprises a heavy chain variable region comprising complementarity determining regions of SEQ ID NOs: 18, 19, and 20 (CDR1, CDR2, and CDR3, respectively) and a light chain variable region comprising complementarity determining regions of SEQ ID

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NOs: 22, 23, and 24 (CDR1, CDR2, and CDR3, respectively) and has immunological reactivity with the CAPRIN-1 protein.

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